

# ***Herpes simplex* virus gD glycoprotein derived peptides as potential drug carriers**

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# Targeted therapy

Cancer therapies:

Surgery  
Radiotherapy

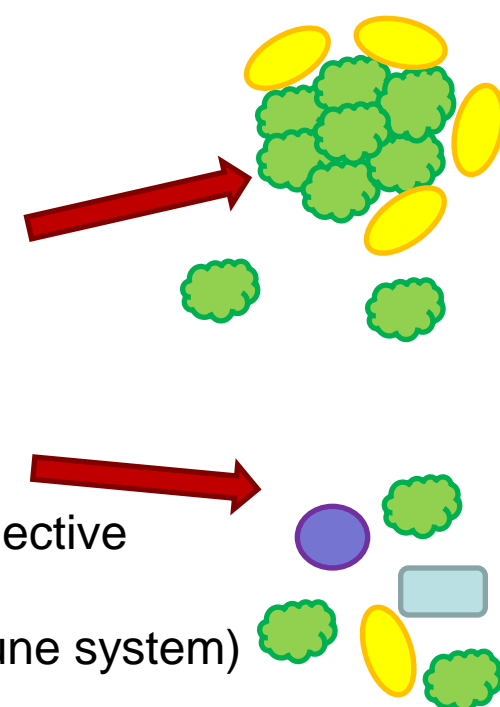
local treatment

Chemotherapy

generally not very selective

Immune therapy (antibodies, boosting own immune system)

Targeted therapy



Drug molecules are specifically targeted to cells with certain features – a special type of cell, a cell with certain receptors on its surface

Inspiration – viruses



# Viruses

Virus: latin, poisonous

Meaning „Agent that causes infectious disease": first recorded in 1728

Louis Pasteur: could not find the causative agent of rabies – is it too small?

Charles Chamberland, 1884: filter with pores smaller than bacteria

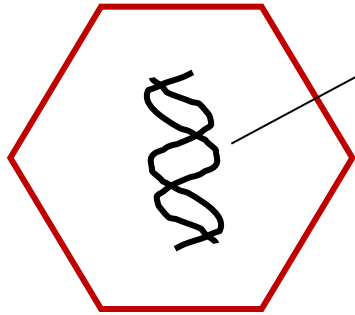
Dimitri Ivanovsky, 1892, tobacco mosaic virus: goes through filter: infectious after filtration – bacterial toxin?

Martinus Beijerinck, 1898, new form of infectious agent in the filtrate „contagium vivum fluidum" (soluble living germ) – re-introducing the word „virus"

Growing viruses first on plant and animal hosts, then on tissue cultures (1906)

Knoll and Ruska, 1931: first electron microscopy image of viruses

# Viruses



DNA or RNA

Single stranded or double stranded  
or ds and ss regions

Linear, circular or segmented

Positive sense, negative sense, ambisense

Protein capsid

*Size:* cca 20 – 500+ nm

*Genome size:* cca 2 000 – 2 000 000 base pairs

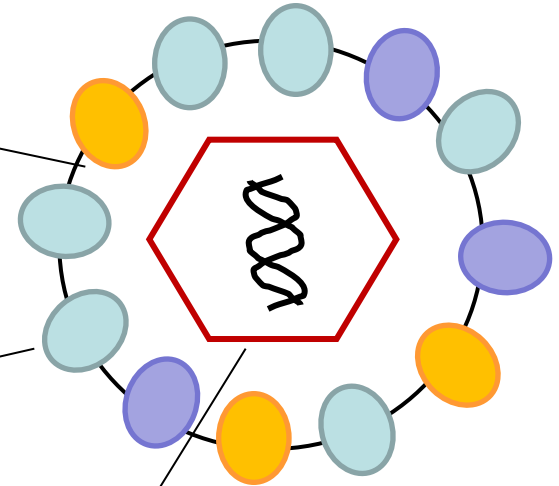
*Shape:* icosahedral, rod-like, spherical, head-tail

Enveloped virus

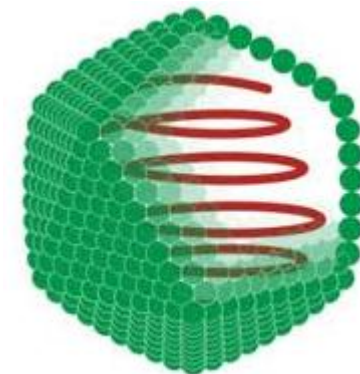
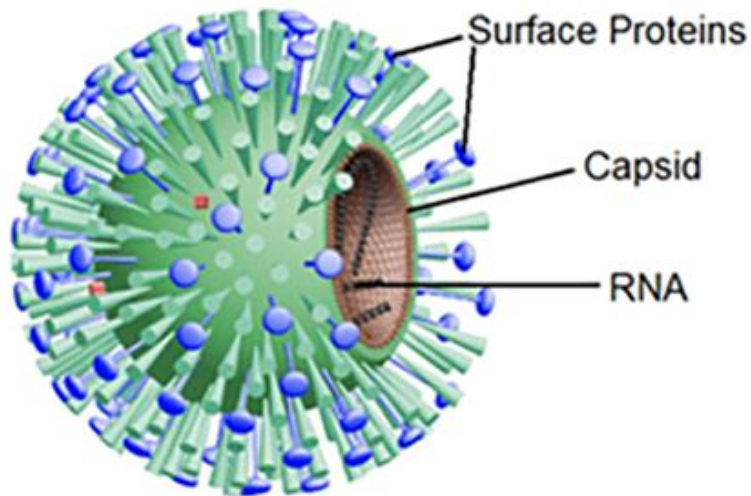
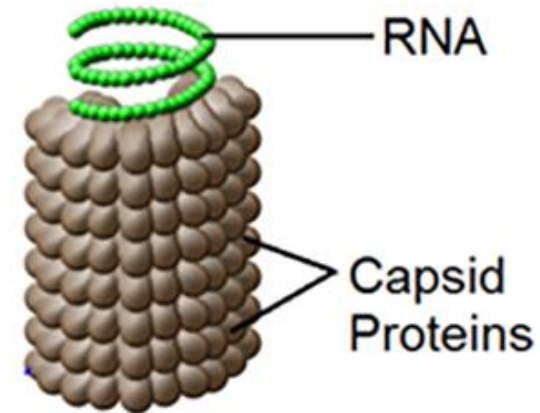
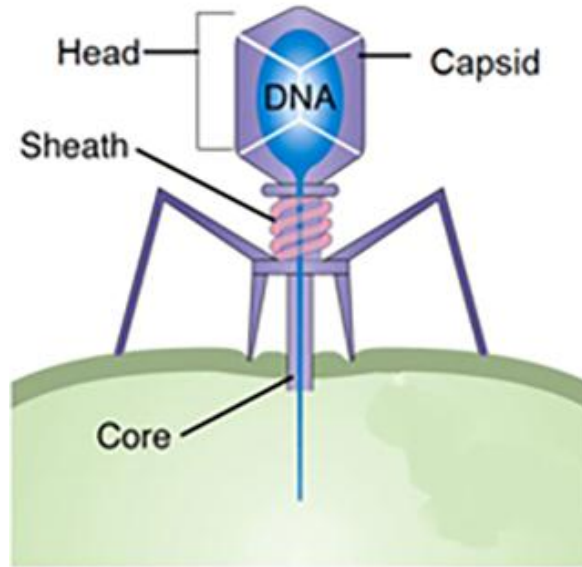
Lipid

Envelope proteins

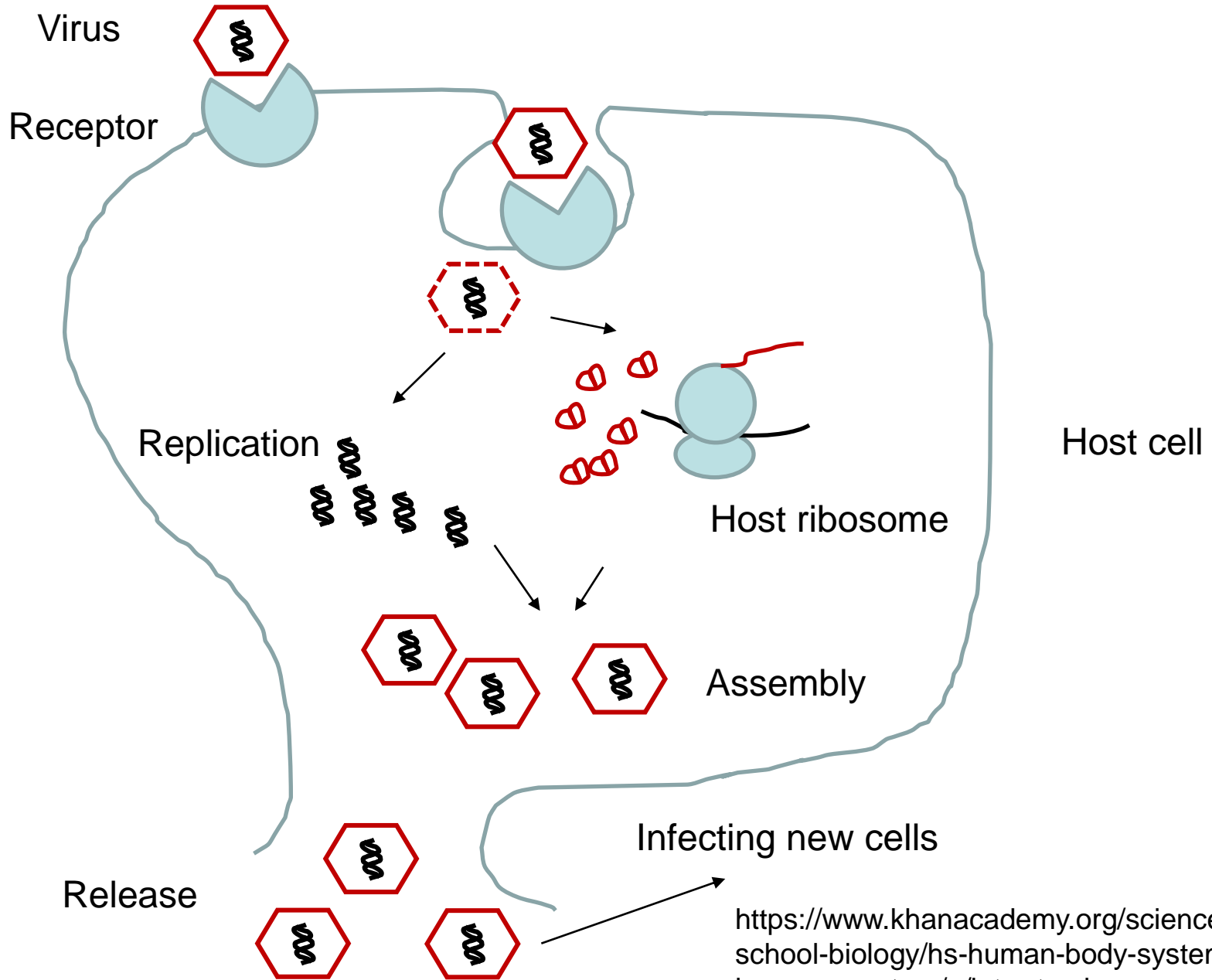
Tegument



# Different shapes viruses



# Virus „life” cycle



<https://www.khanacademy.org/science/high-school-biology/hs-human-body-systems/hs-the-immune-system/a/intro-to-viruses>

## **Baltimore classification** (David Baltimore, Nobel prize)

I: dsDNA viruses (e.g. Adenoviruses, Herpesviruses, Poxviruses)

II: ssDNA viruses (+ strand or "sense") DNA (e.g. Parvoviruses)

III: dsRNA viruses (e.g. Reoviruses)

IV: (+)ssRNA viruses (+ strand or sense) RNA (e.g. Picornaviruses, Togaviruses)

V: (-)ssRNA viruses (- strand or antisense) RNA (e.g. Orthomyxoviruses, Rhabdoviruses)

VI: ssRNA-RT viruses (+ strand or sense) RNA with DNA intermediate in life-cycle (e.g. Retroviruses)

VII: dsDNA-RT viruses DNA with RNA intermediate in life-cycle (e.g. Hepadnaviruses)

## **International Committee on Taxonomy of Viruses (ICTV) classification**

Order (-virales)

Family (-viridae)

Subfamily (-virinae)

Genus (-virus)

Species (-virus)

# Herpesvirales (order), Herpesviridae (family)

More than 130 species, 9 infecting humans (HHV)

Subfamilies:

## $\alpha$ – herpesvirinae

HSV-1, HSV-2, VZV  
HHV-1, HHV-2, HHV-3

## $\beta$ – herpesvirinae

HCMV, HHV-6A, HHV-6B, HHV-7  
HHV-5,

## $\gamma$ – herpesvirinae

EBV, KSHV  
HHV-4, HHV-8

HSV: *Herpes simplex* virus (genus)

VZV: *Varicella zoster* virus, chickenpox

HCMV: Human cytomegalovirus

EBV: Epstein-Barr virus

KSHV: Kaposi's Sarcoma associated Herpes virus



# *Herpes simplex virus 1 (HSV-1, HHV-1)*

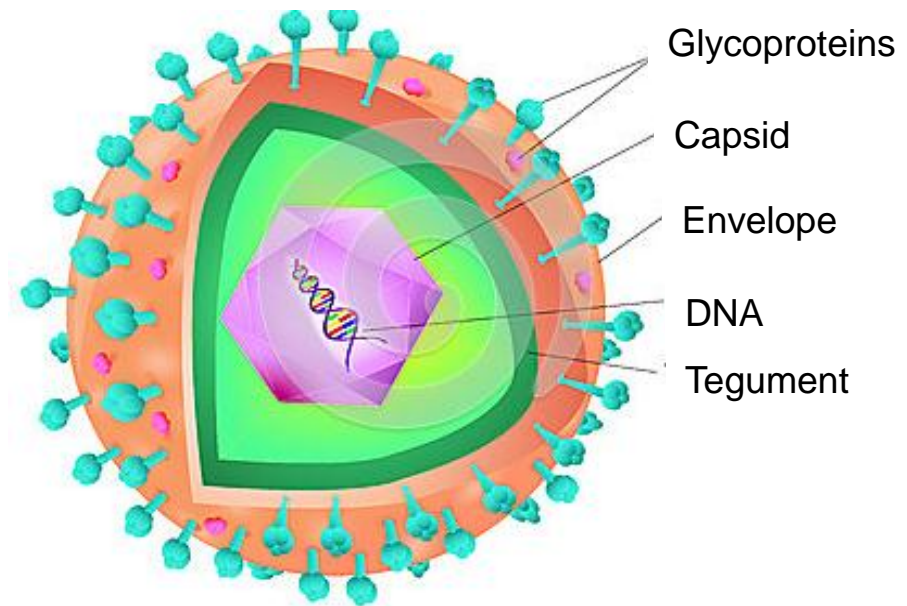
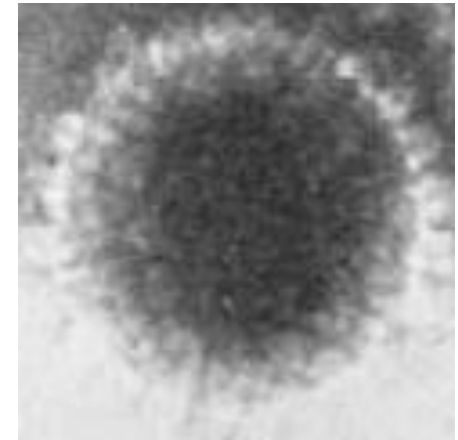
Herpesviridae,  $\alpha$ -Herpesvirinae

Labial herpes (cold sores)

Complications:  
meningitis, encephalitis  
blindness

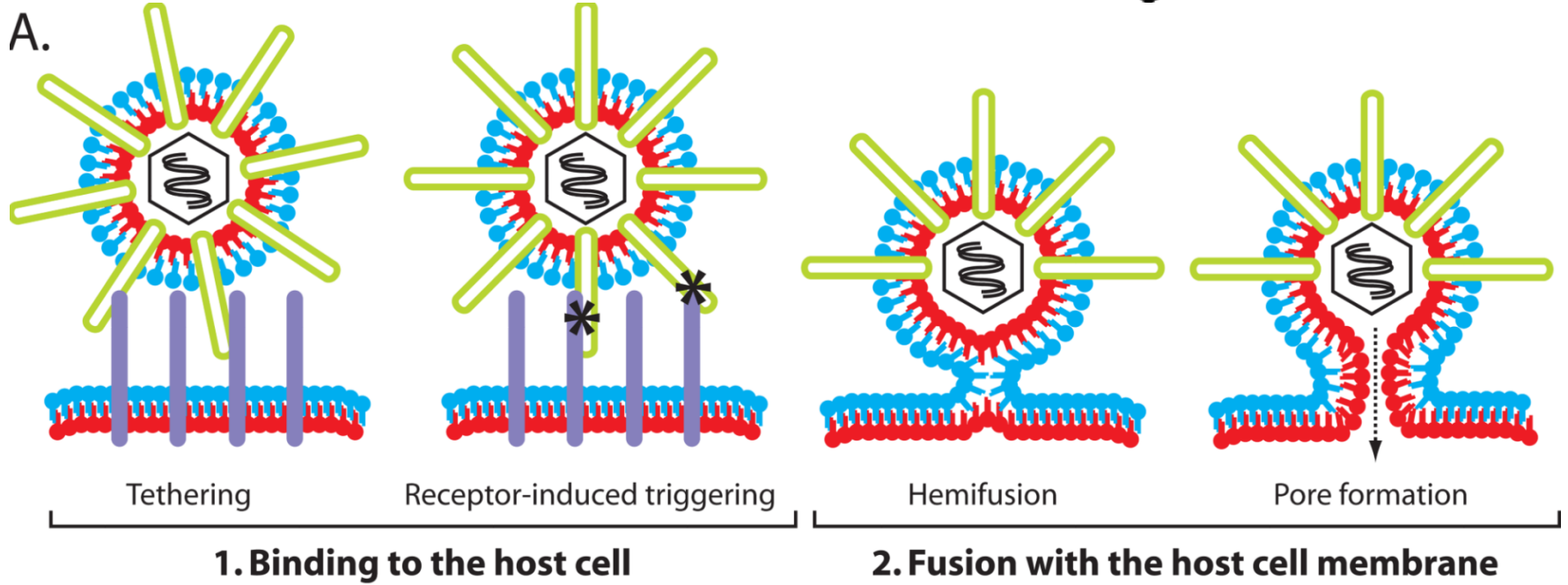
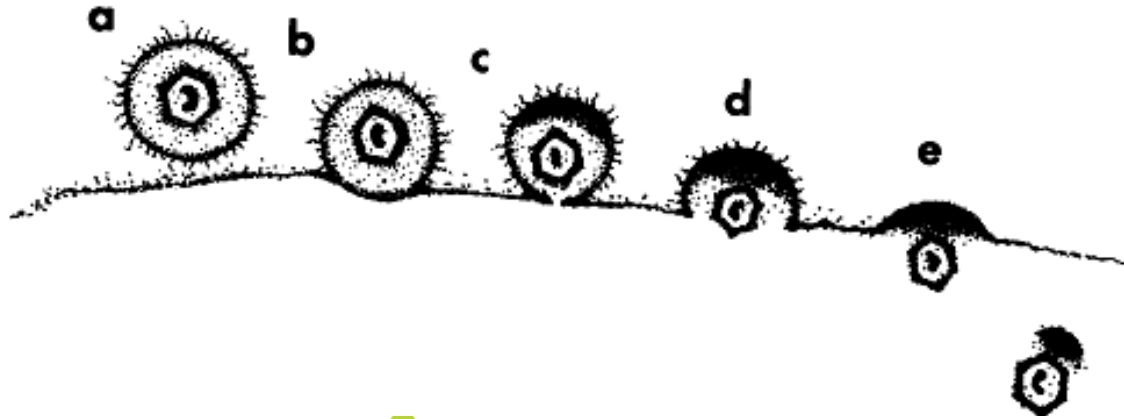
Frequent recurrence,  
latency, in neurons  
Evading the immune system

Treatment  
Nucleoside derivatives  
DNA inhibitors

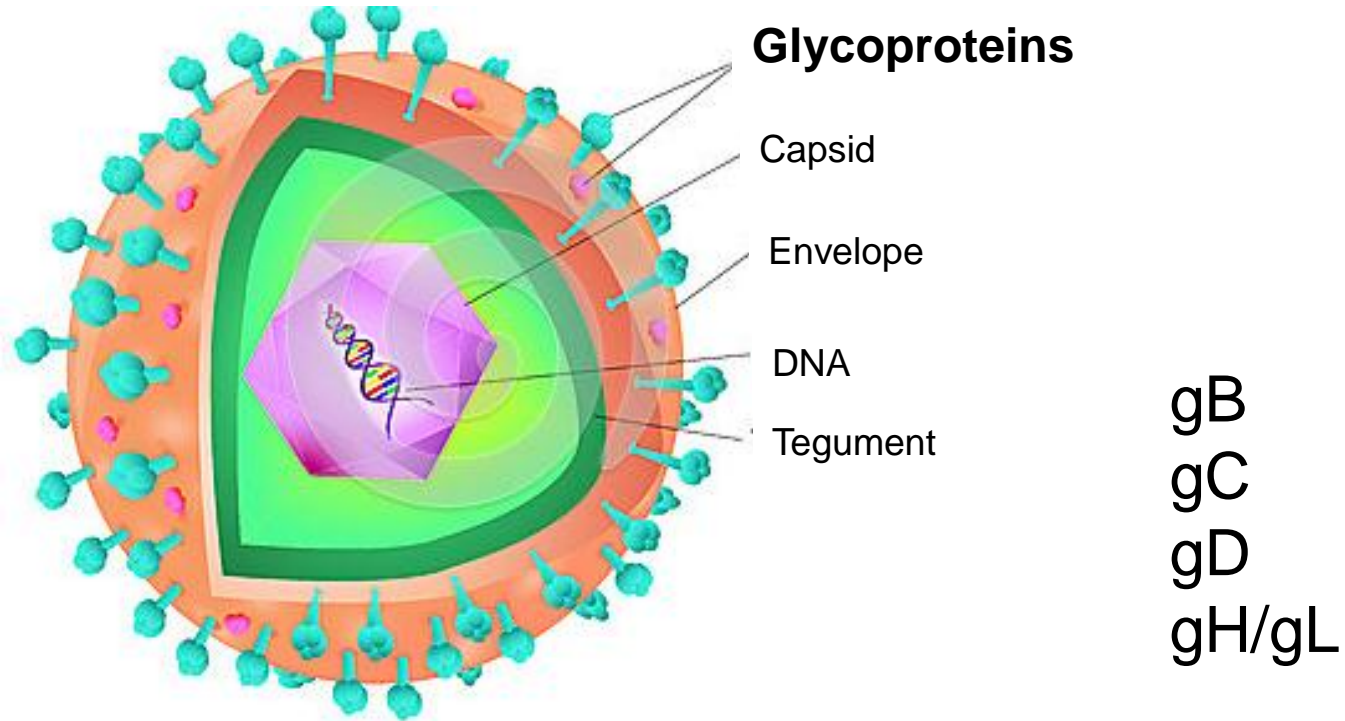


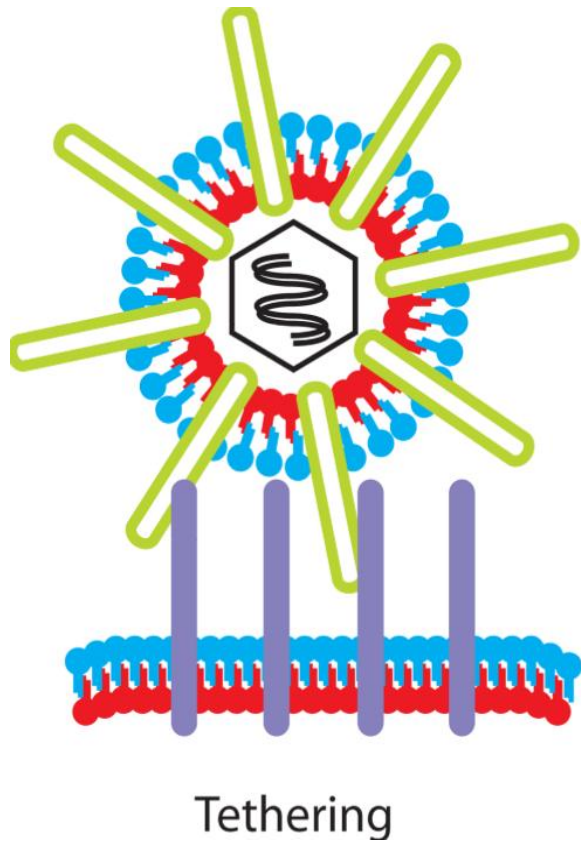
125 nm, 153 000 bp, ds DNA,  
icosahedral capsid

# Cellular entry of *Herpes simplex virus*



# Participating glycoproteins in HSV entry

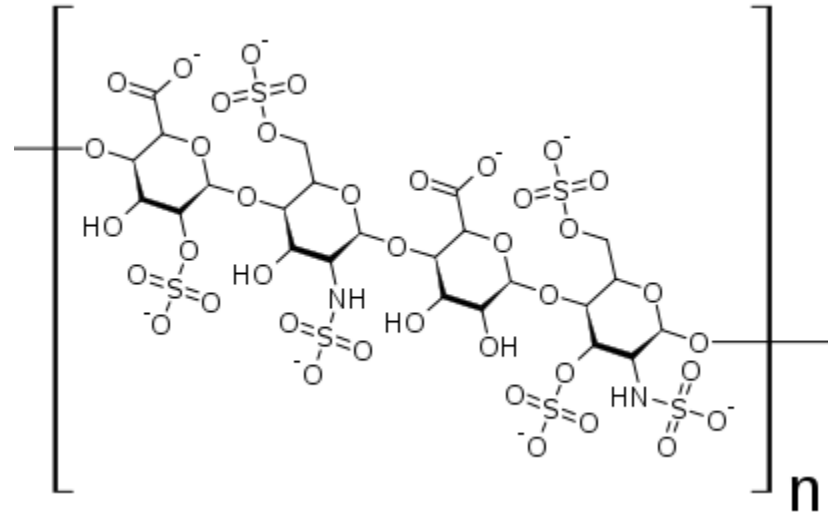


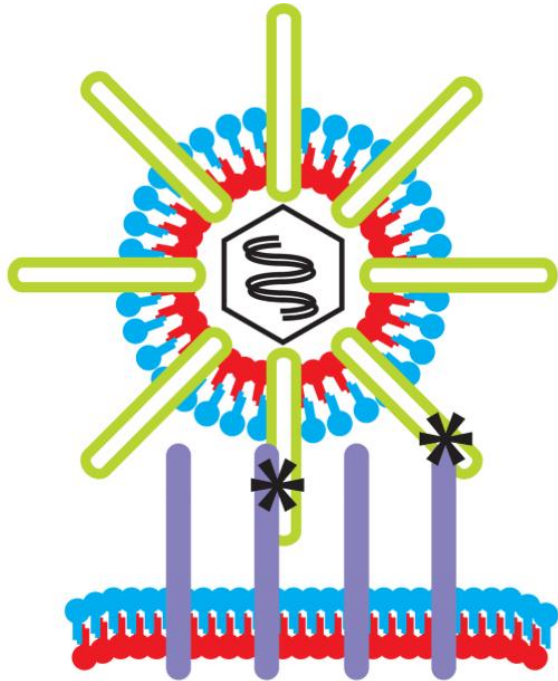


## Tethering

Concentrating viruses on the cell surface  
Does not specifically trigger fusion

gB and gC interact with heparan sulfate





Receptor-induced triggering

## Receptor induced triggering

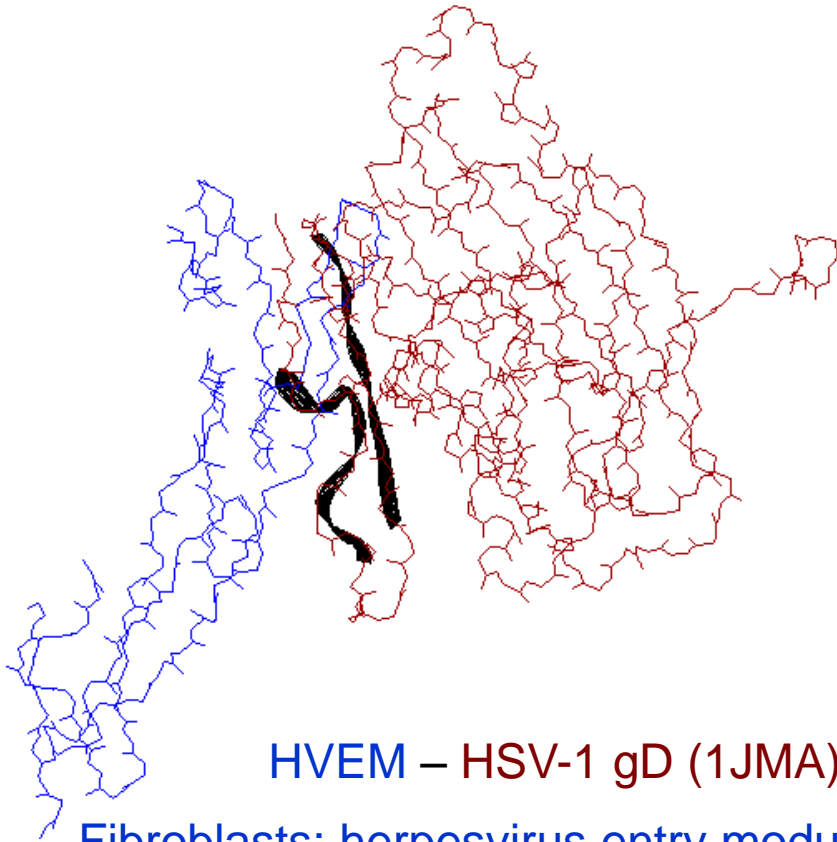
gD is the main receptor binding glycoprotein, unique for *Herpes simplex* 1 and 2 viruses

Receptors:

***HVEM*** (herpes virus entry modulator, immunomodulator, tumor necrosis factor receptor superfamily)

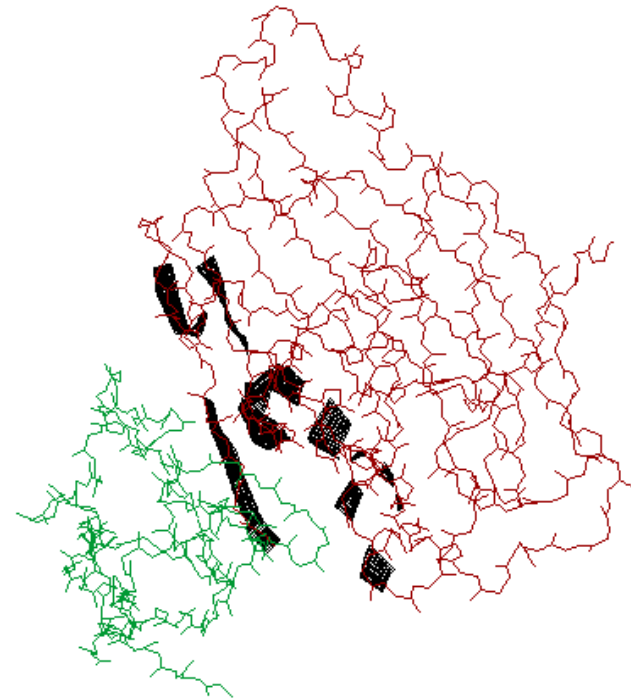
***Nectin-1/2*** (cell adhesion molecules)

# Binding of **HSV gD** to **HVEM** and **nectin** receptors



**HVEM – HSV-1 gD (1JMA)**

Fibroblasts: herpesvirus entry modulator A (HVEM), immunomodulator, tumor necrosis factor receptor superfamily.



**Nectin – HSV-1 gD (3U82)**

Neurons, keratinocytes, epithelial cells: nectin-1 adhesion protein

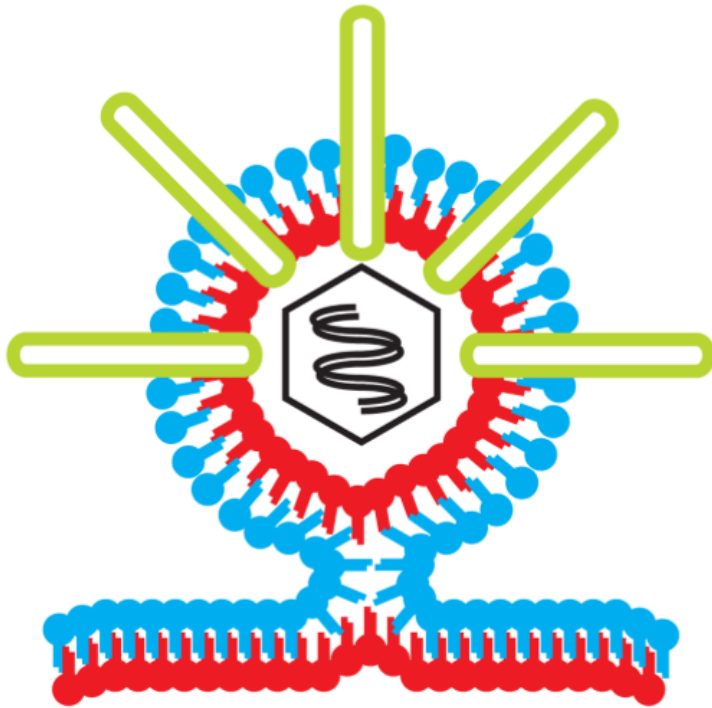
# Hemifusion

Fusogenic glycoproteins

gB, **gH/gL**

(in other herpes viruses as well,  
e.g. Epstein-Barr)

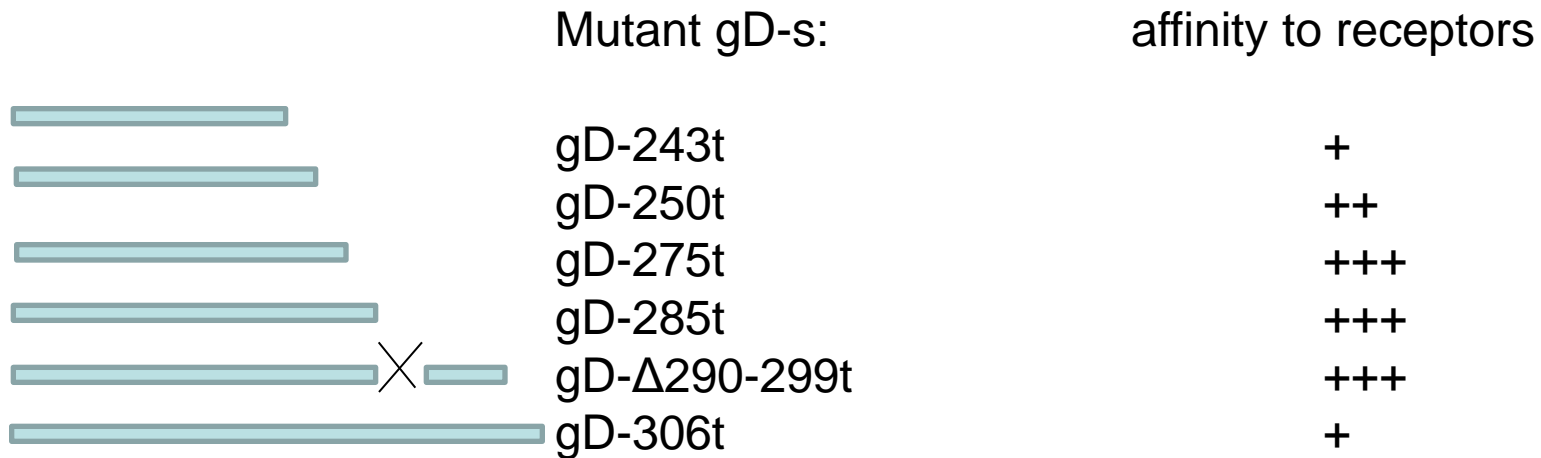
gH has **amphiphil helices**,  
effecting pore formation



Hemifusion



# Structural requirements of gD for infection



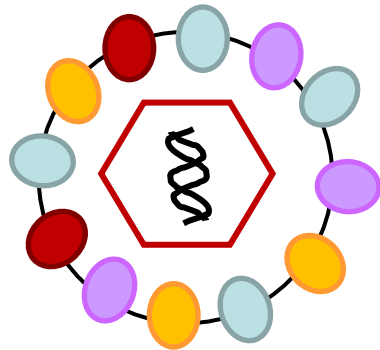
Milne et al, 2003, J Virol 77, 8962-8972

Krummenacher et al, 1998, J. Virol 72, 7064-7074

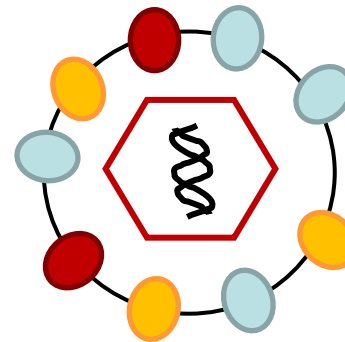


# Structural requirements for gD for infection

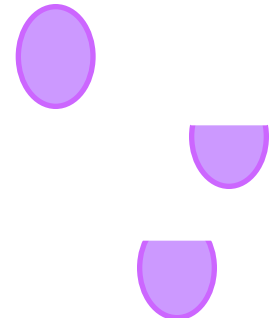
Can soluble gD substitute the virion-bound gD?



HSV wt



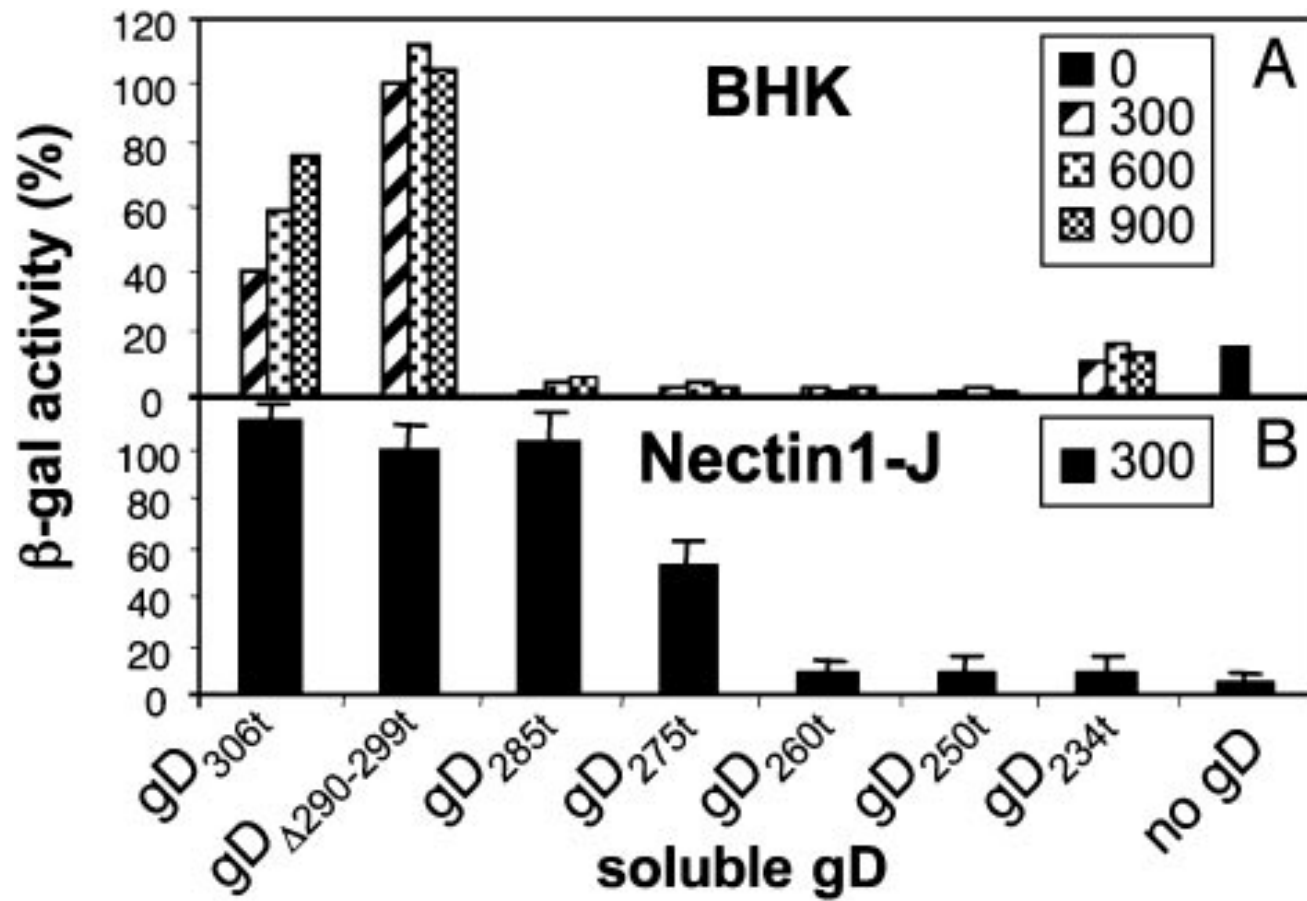
HSV gD null



gB  
gC  
gD  
gH/gL

	BHK cells	Nectin 1-J cells
gD-243t	-	-
gD-250t	-	-
gD-260t	-	-
gD-275t	-	+
gD-285t	-	+++
gD- $\Delta$ 290-299t	+++	+++
gD-306t	++	+++

(hamster kidney, fibroblast, nectin expressing cells)



# Structural requirements for gD for infection

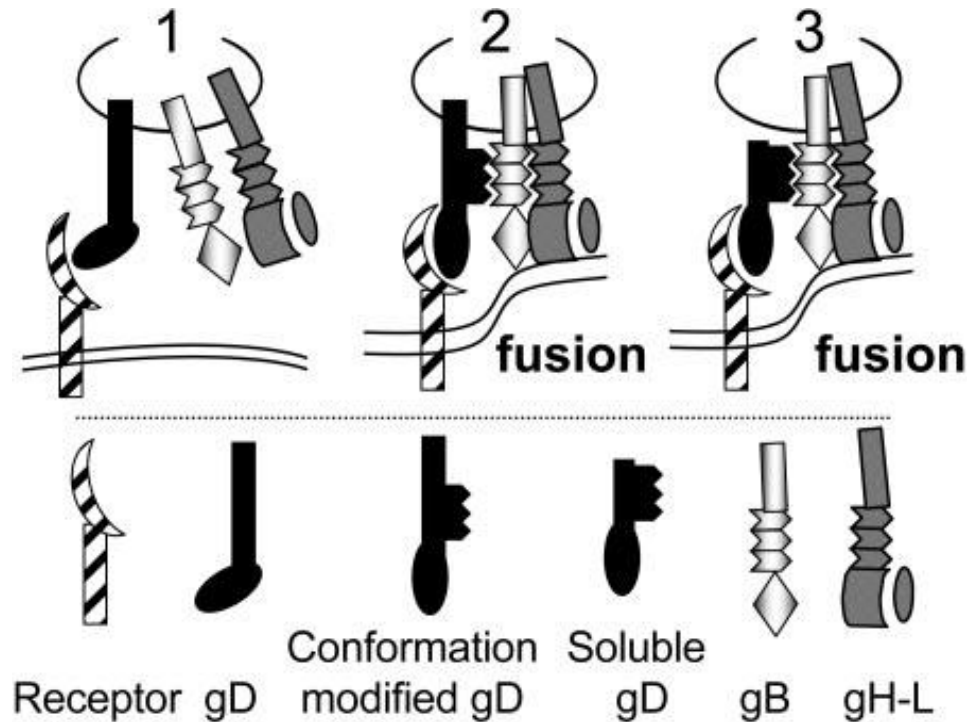
Receptor binding domain

Pro-fusion domain  
260-307

TM, C-term:  
Not needed for infection

Role of gD:  
Not only receptor binding!

to form a tripartite complex  
Switch of fusion glycoproteins to  
fusion-active state



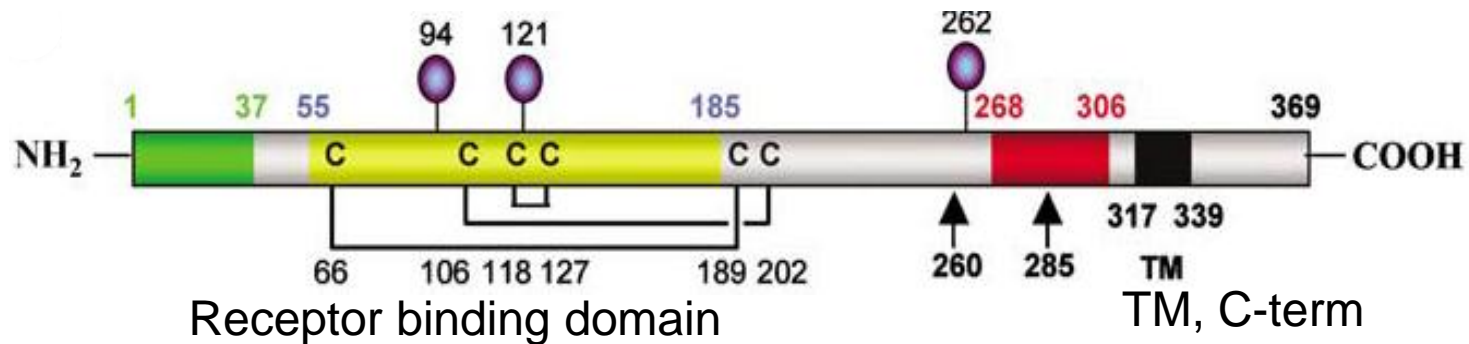
2005: three X-ray structures,

gD 1-285

gD 1-316

gD 1-316 bound to HVEM

260-285 disordered, not visible in X-ray  
C-terminal not resolved

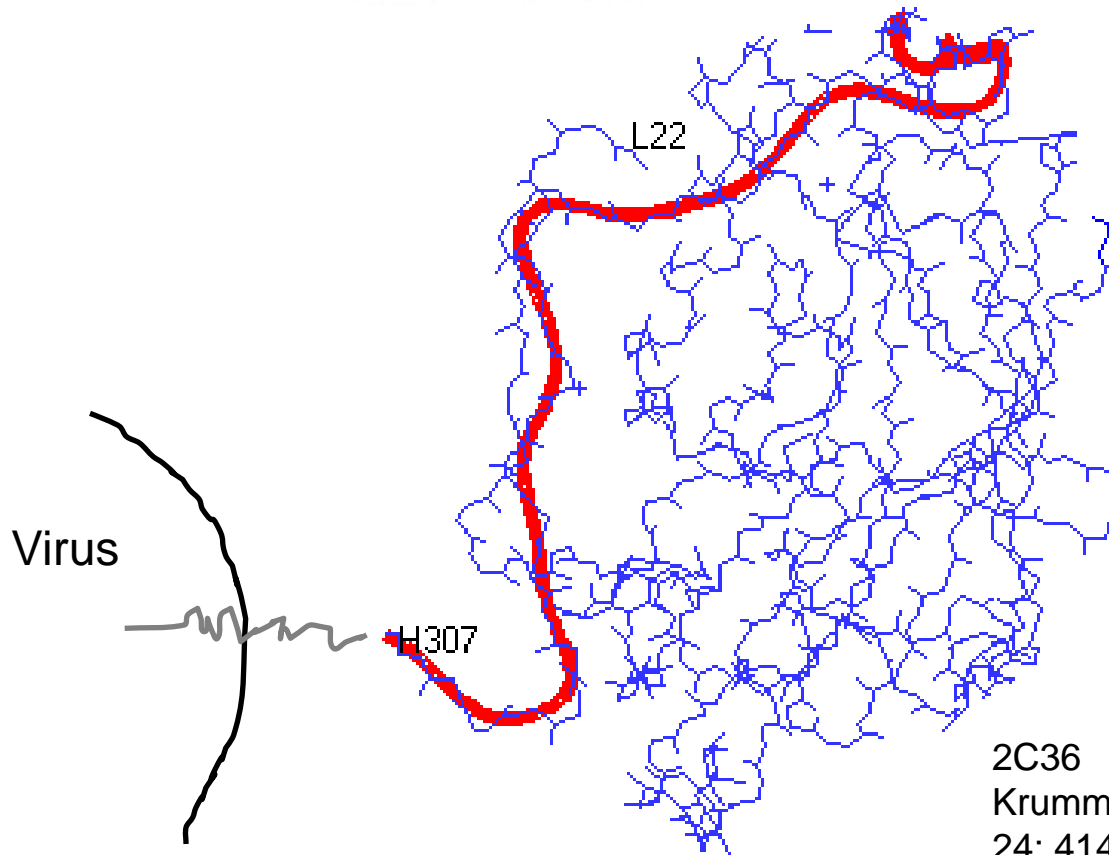
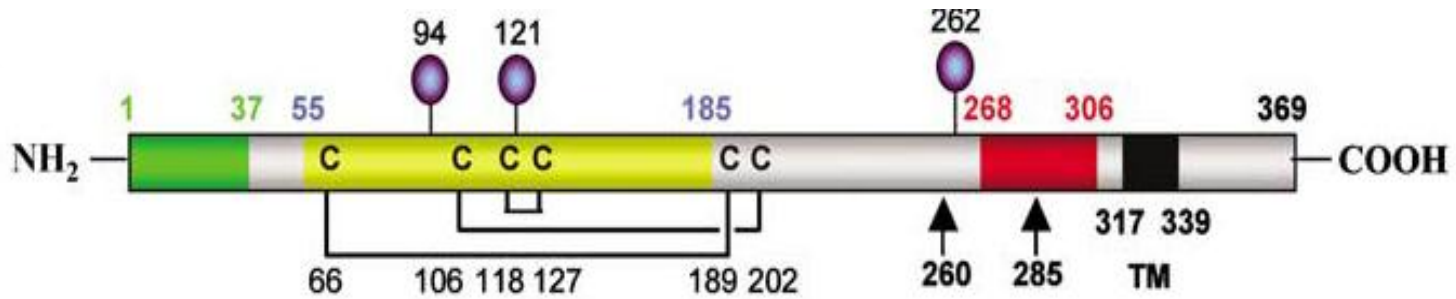


Pro-fusion domain  
260-307

New: gD 1-306, + mutant  
pre-receptor binding conformer

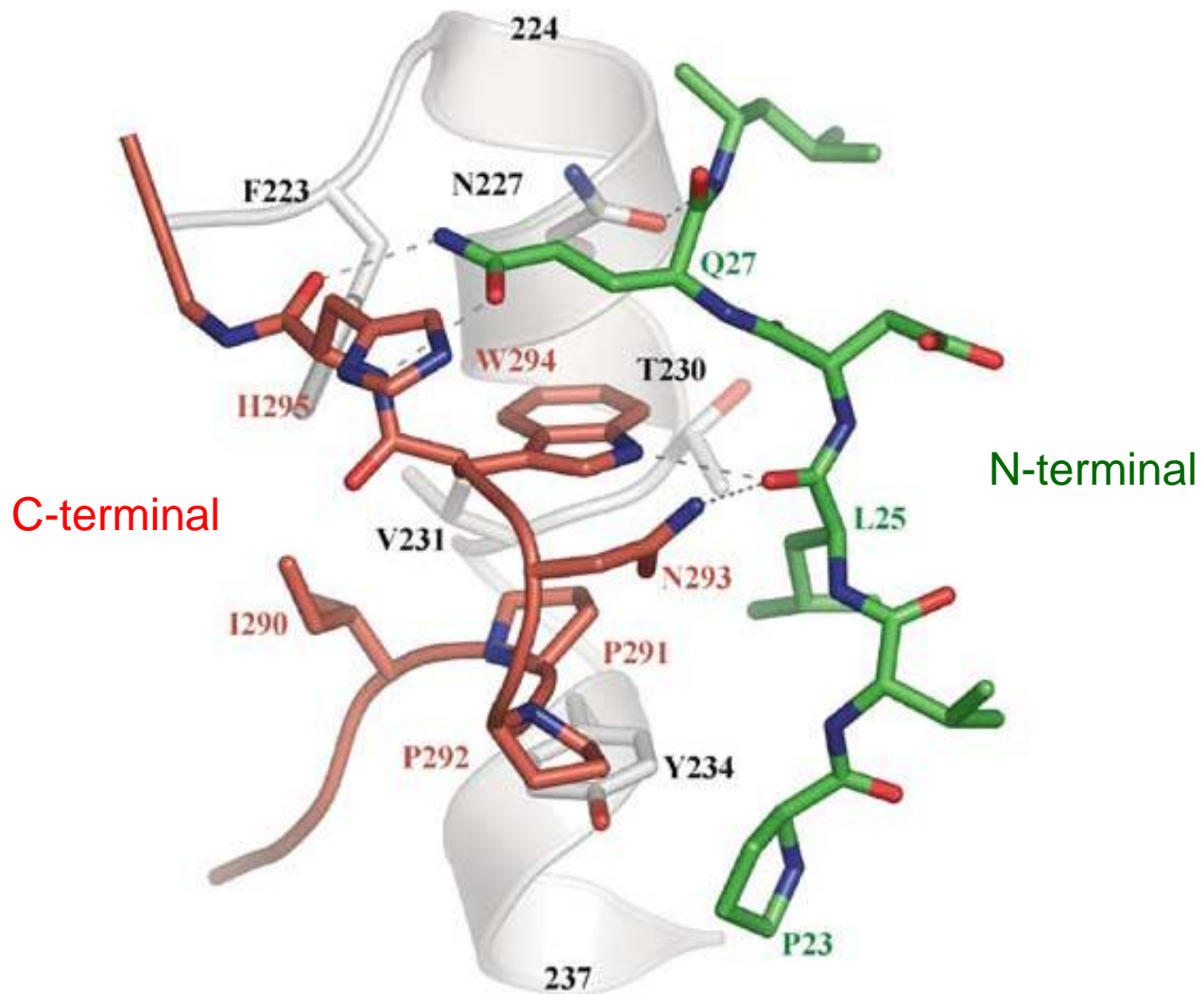
(N-terminal not resolved)

# The structure of HSV-1 gD glycoprotein



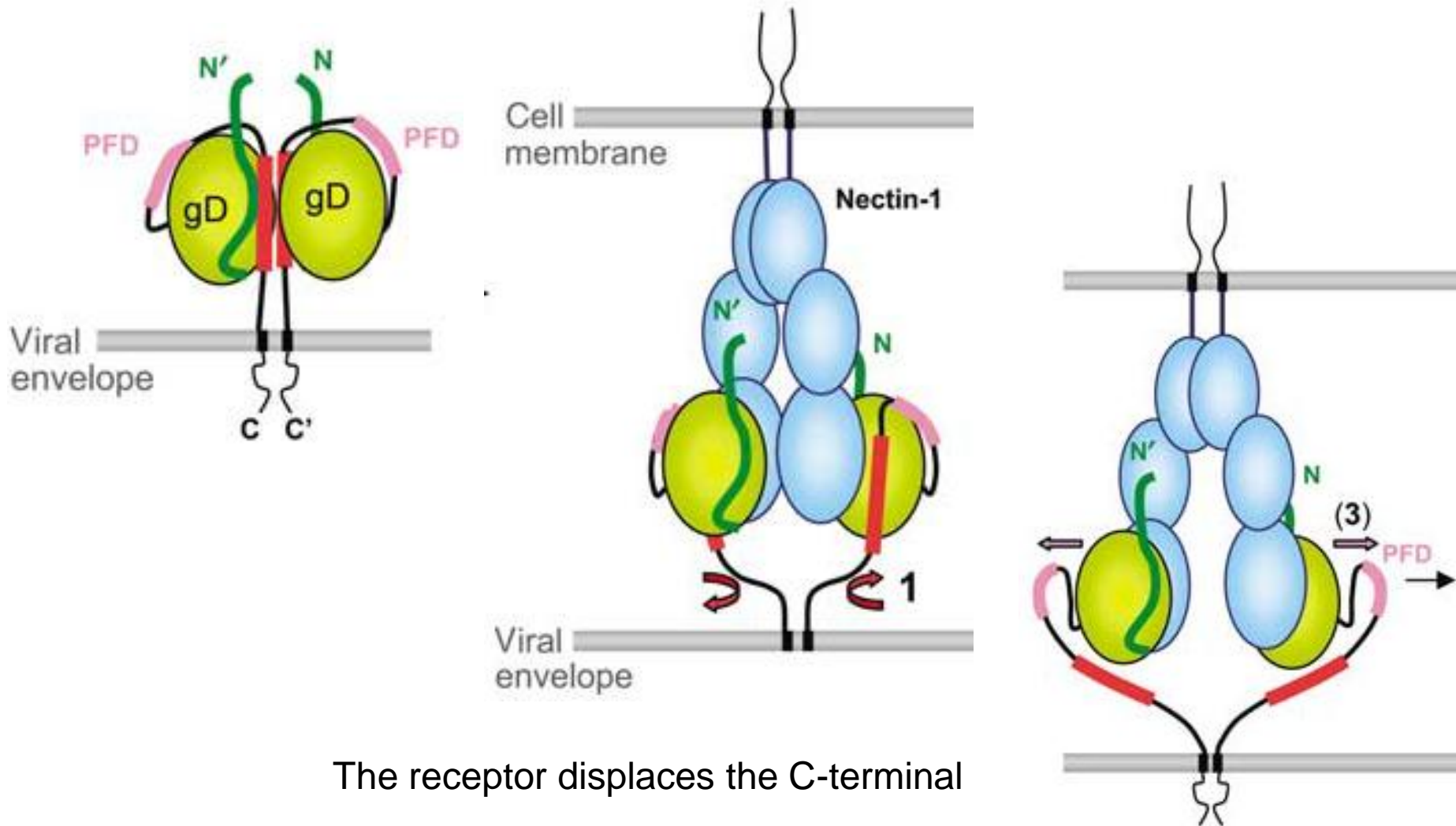
2C36

Krummenacher et al: EMBO J,  
24: 4144-4153 (2005)



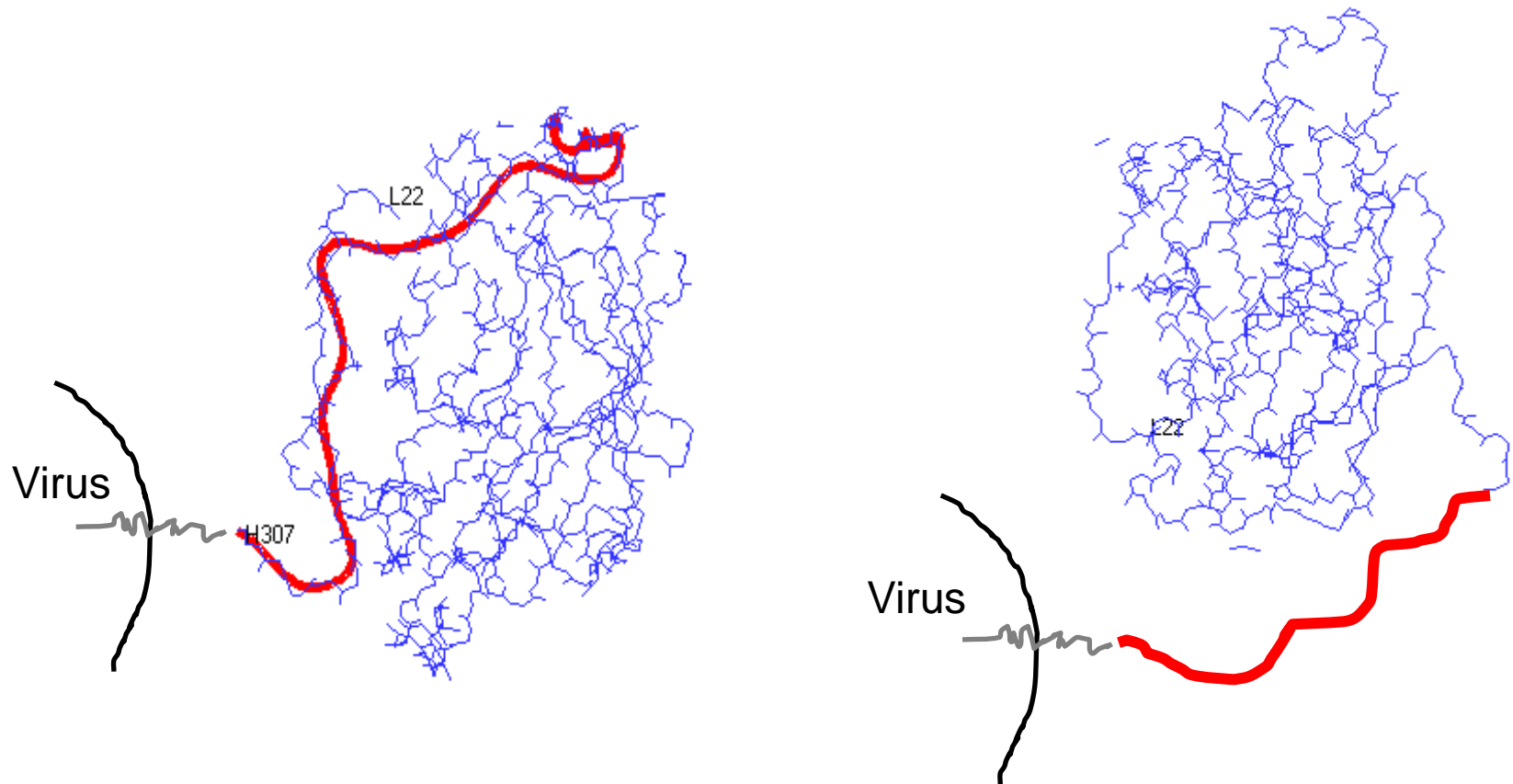
Flexible C-terminal,  
Conformational equilibrium

# Conformational changes during receptor binding of HSV gD



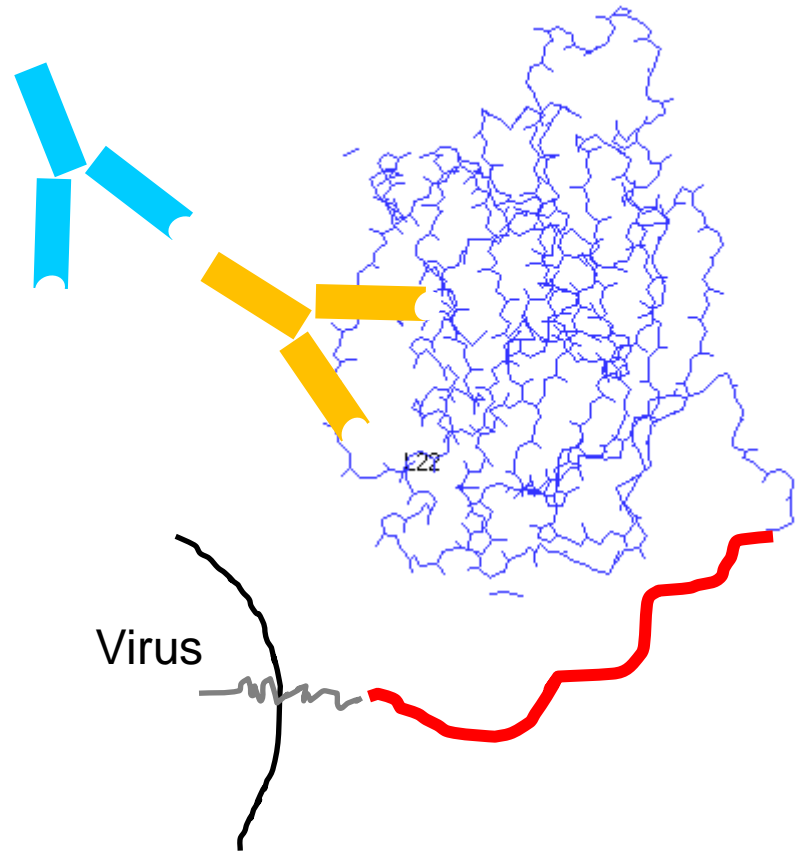
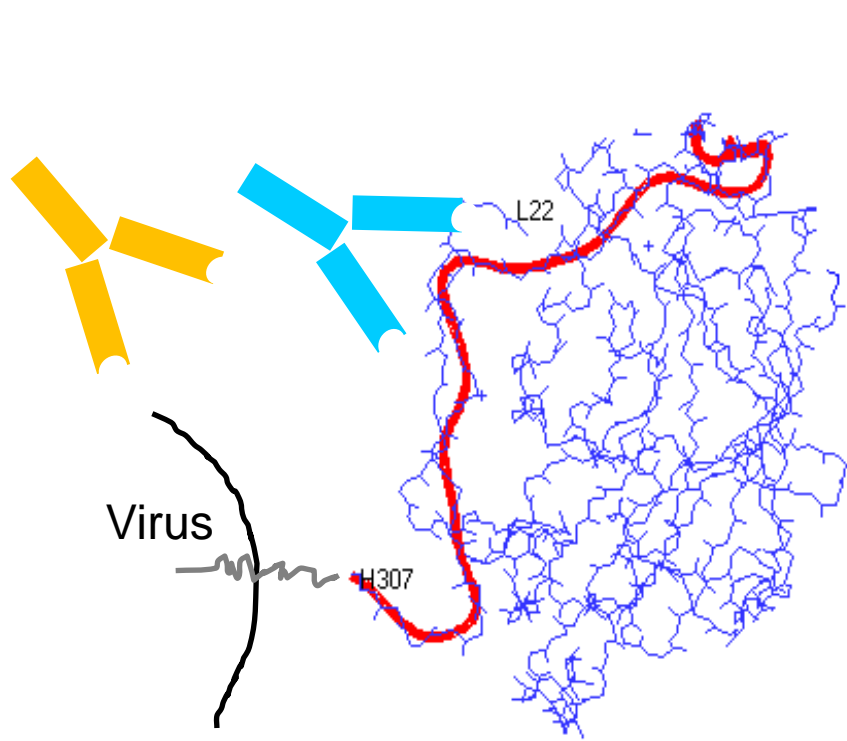
The receptor displaces the C-terminal

# Conformational changes during receptor binding of HSV gD



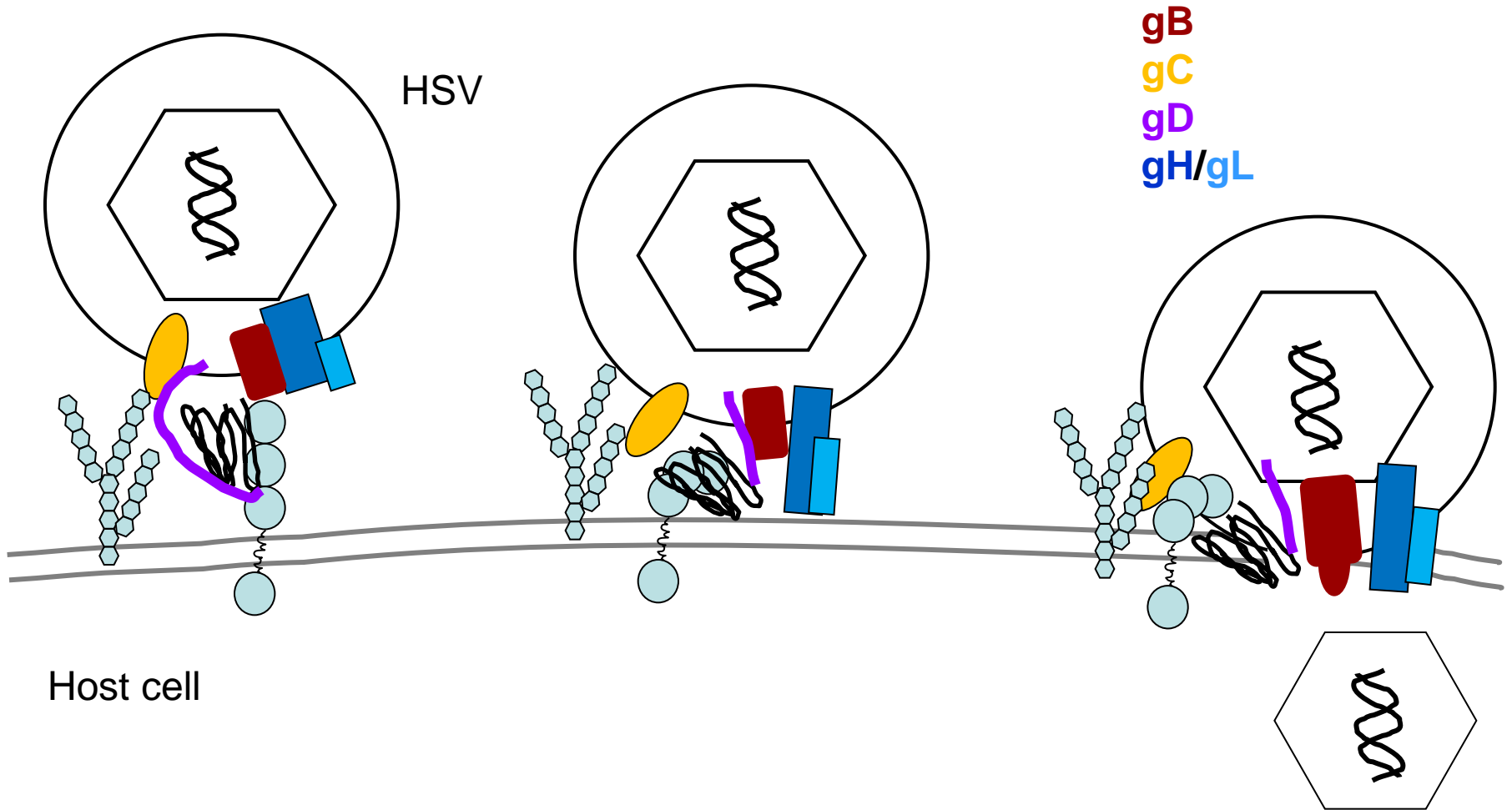
Masking both nectin and HVEM binding sites  
Possibly gB and gH/gL binding sites are blocked



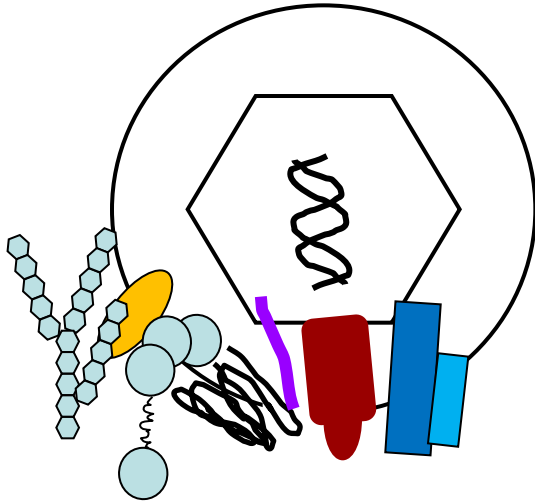


Protection against the host's immune system

# Cellular entry of *Herpes simplex virus*



# Using virus peptides for cellular targeting

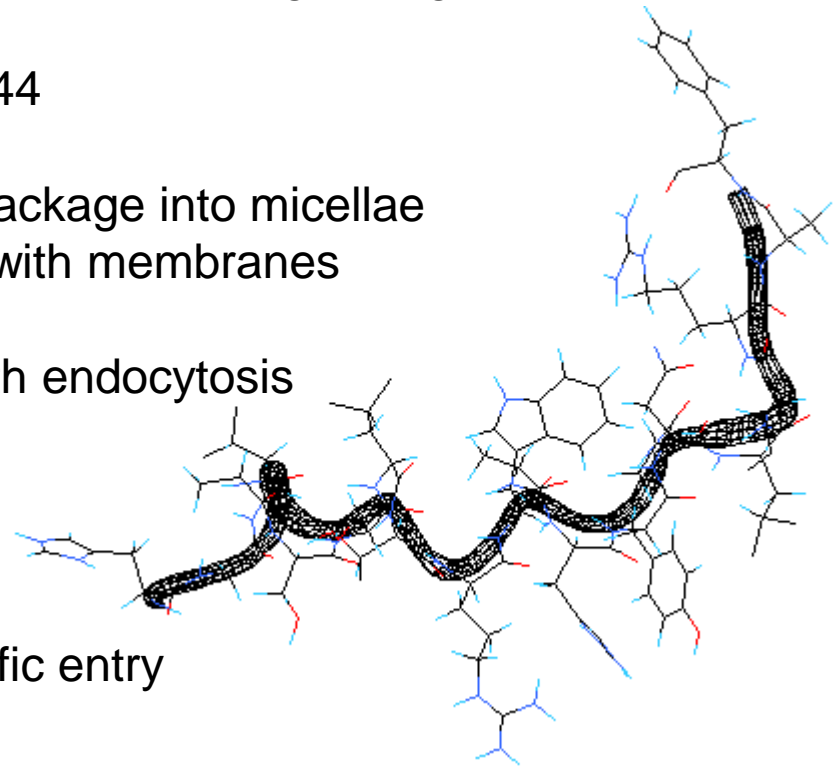


**gH** derived pore forming, fusogenic peptide

gH 625-644

Easy to package into micellae  
Interacts with membranes

Enters with endocytosis



**Not** specific entry

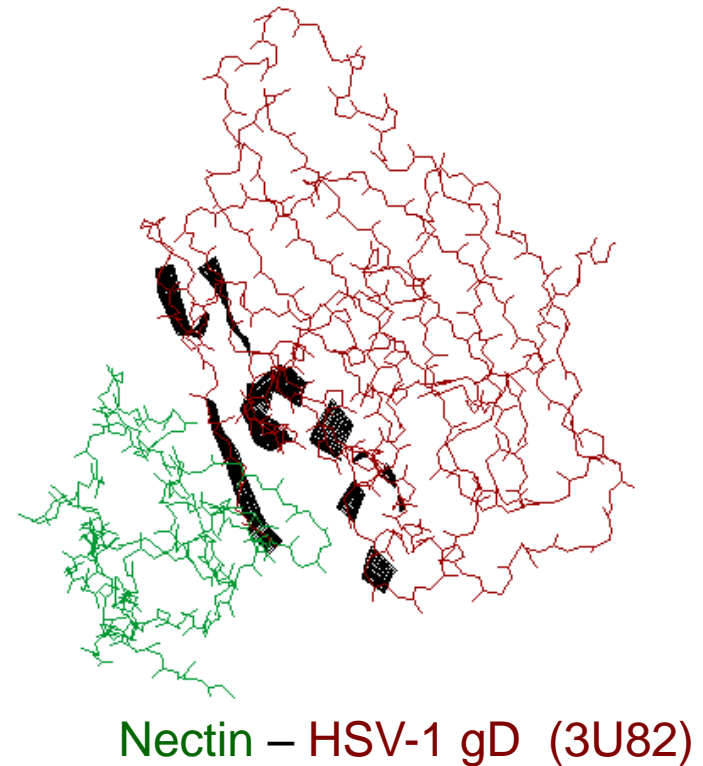
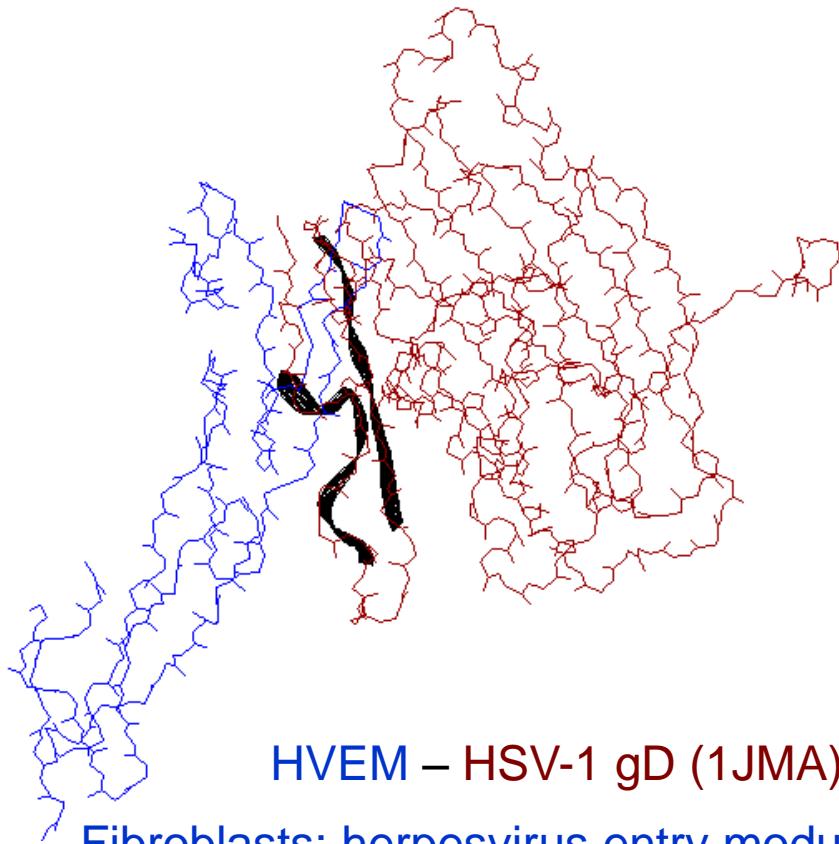
Galdiero et al, Biochemistry. 2012;51(14):3121-3128.  
Smaldone et al, Int J Nanomed 2013;8 2555–2565

## Our aims:

Finding synthetic peptides effectively and specifically internalising into cells carrying nectin or HVEM receptor, with receptor mediated cellular uptake

Peptide sequences based on the receptor binding of HSV-1 gD

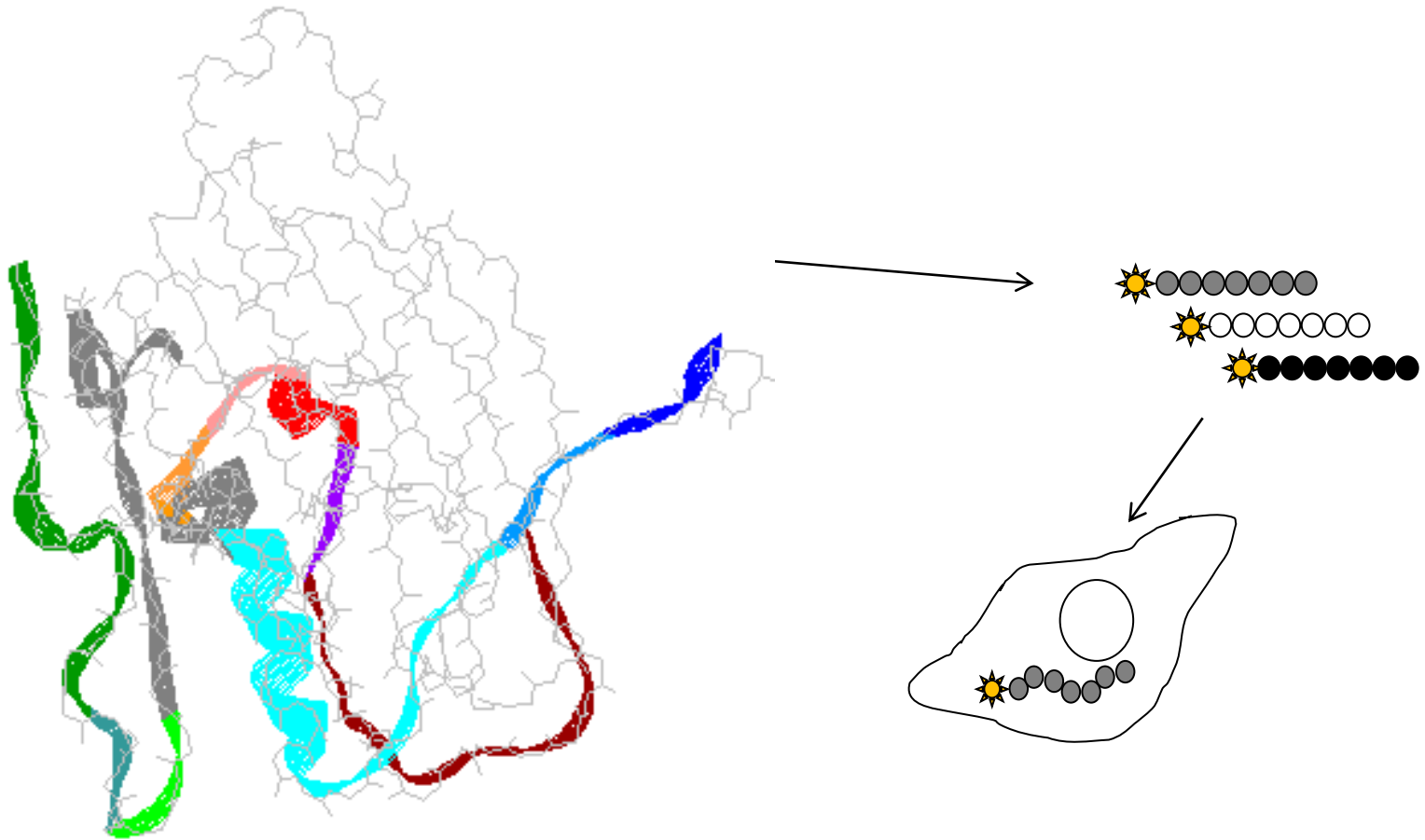
# Binding of HSV gD to HVEM and nectin receptors



Fibroblasts: herpesvirus entry modulator A (HVEM), immunomodulator, tumor necrosis factor receptor superfamily.

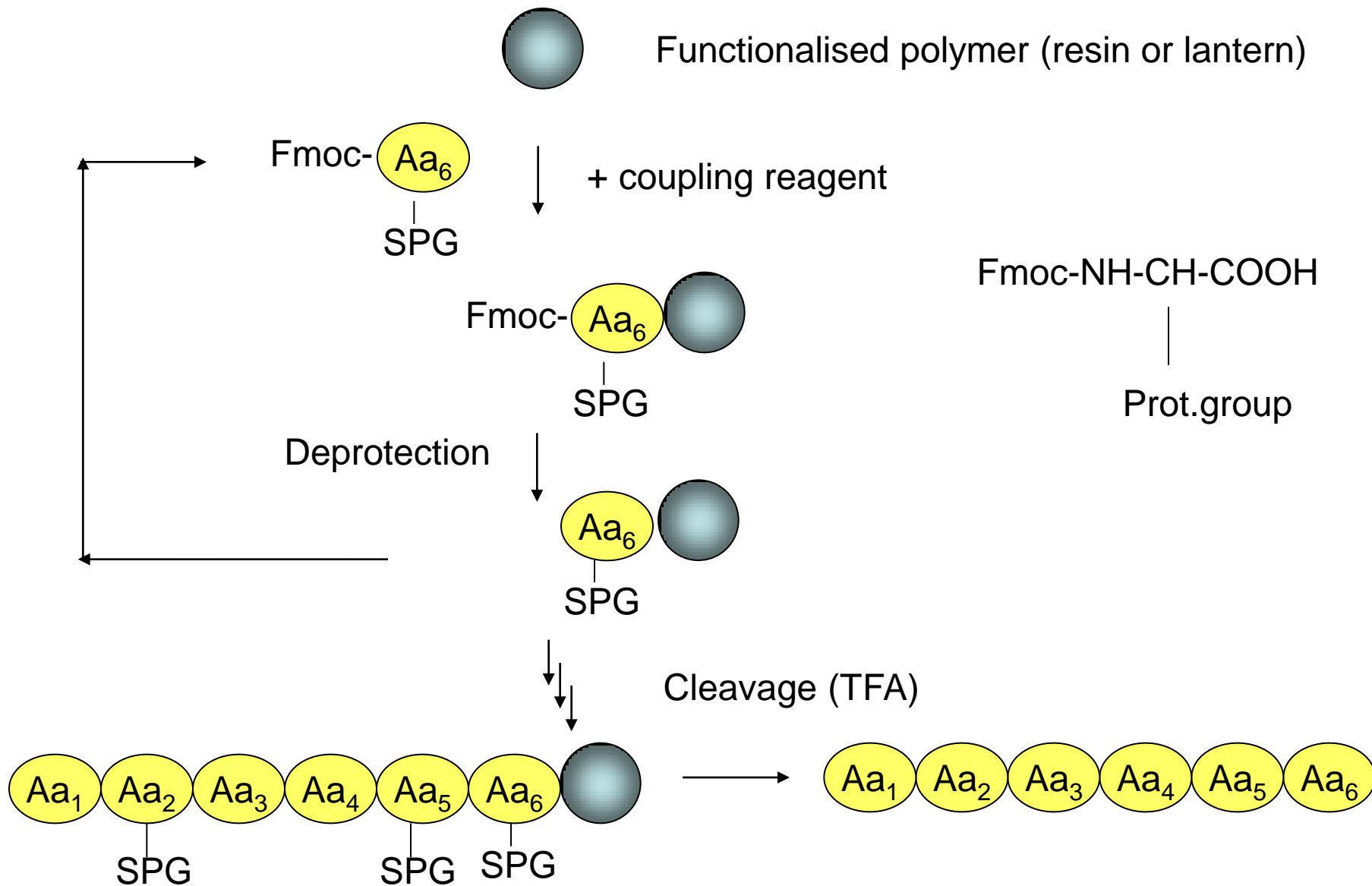
Neurons, keratinocytes, epithelial cells: nectin-1 adhesion protein

# Strategy



21 **HGVRGKYALADASLKMADPNRFRGKDL**PVLDQLTDPPGVRRVYHIQA<sup>67</sup>  
206 **LEHRAKGSCKYALPLRIPPSACLSPQAYQQGVT**DS<sup>241</sup>  
239 **VDSIGMLPRFIPENQRTVAVYSLKIAGWHGPKAPYTSTLL**PP<sup>280</sup>

# Solid phase peptide synthesis scheme



# Peptide synthesis

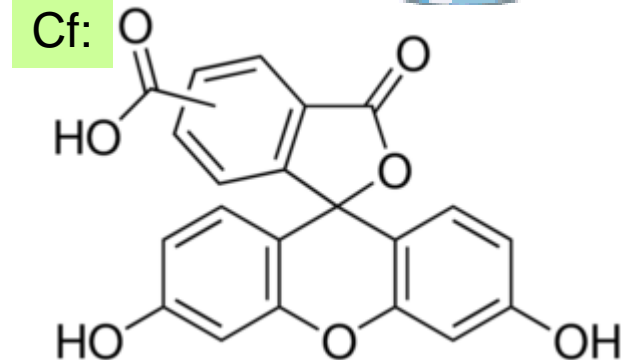
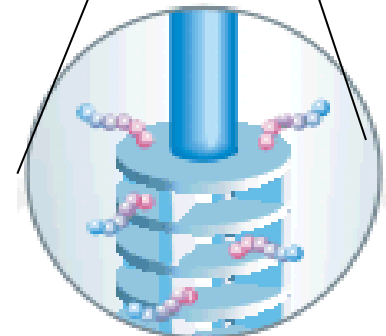
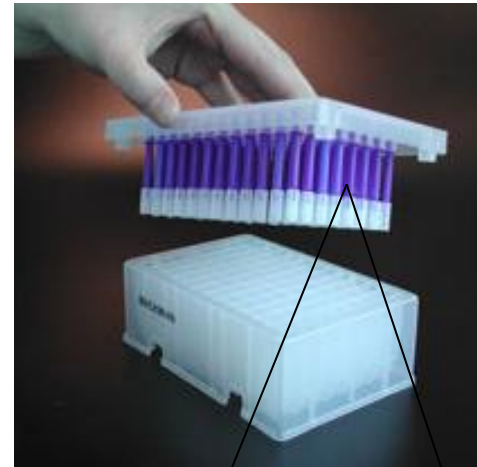
Carrier: SynPhase™ lantern (Mimotopes),  
Capacity: 8 μmol  
Fmoc/tBu strategy

Fmoc cleavage: piperidine – DBU – DMF  
(2:2:96 V/V/V)

DIC/HOBt coupling 2x, 10 eq  
Monitoring: bromophenolblue

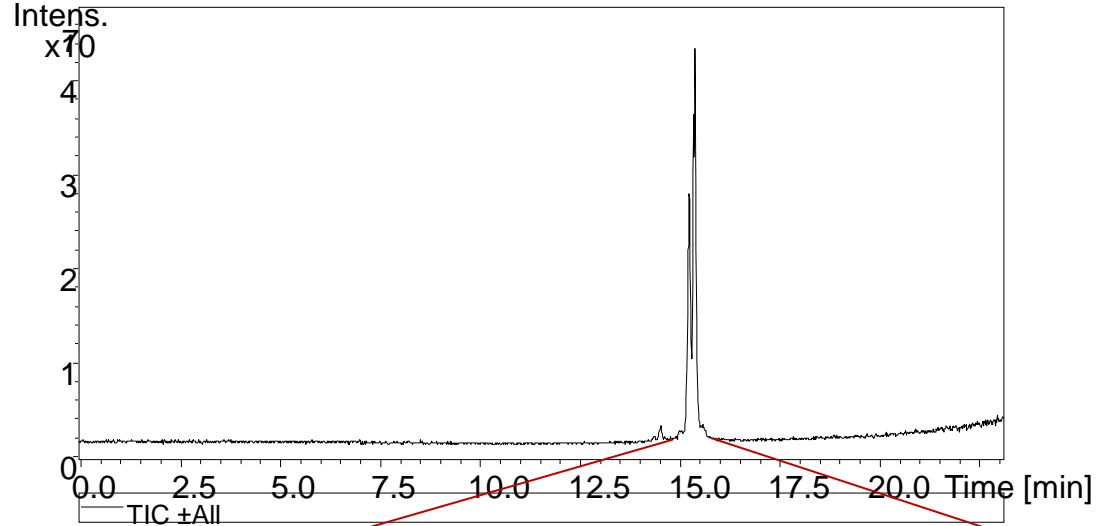
Labelling: 5(6)-carboxyfluorescein (Cf),  
DIC/HOBt coupling

Cleavage: 1.5 h  
TFA – thioanisol – water – phenol –  
1,2-ethandithiol (80:5:5:7.5:2.5, V/V/V/m/V)

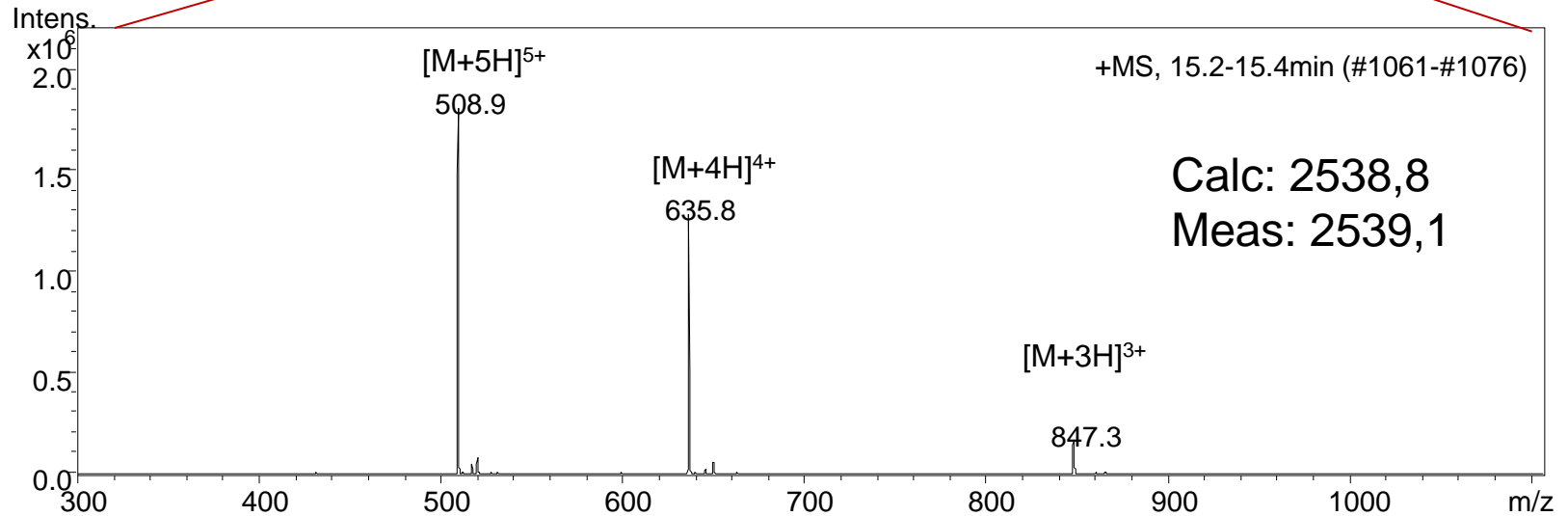




# HPLC-MS chromatogram of a purified Cf-HSV-gD peptide

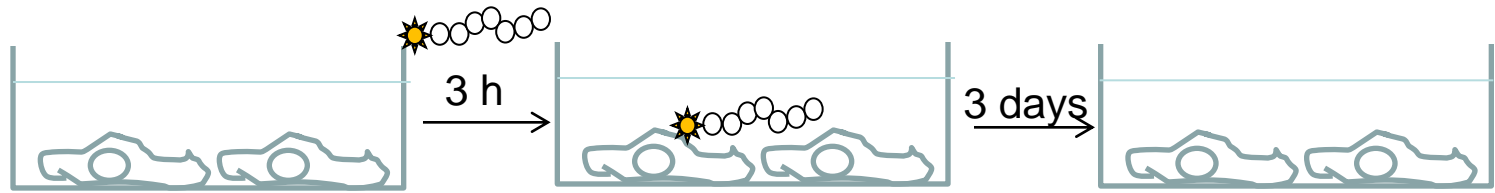


Jasco PU-2085 Plus Semi-Micro HPLC, Phenomenex Synergi C18, 100 x 2,0 mm, 2.5  $\mu\text{m}$ , 100  $\text{\AA}$



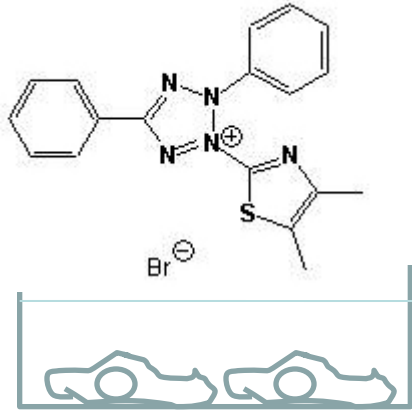
Bruker Daltonics Esquire 3000+

# *In vitro* cytostatic effect of Cf-HSV peptides, MTT assay



3-5x10<sup>3</sup> SH-SY5Y  
neuroblastoma cells

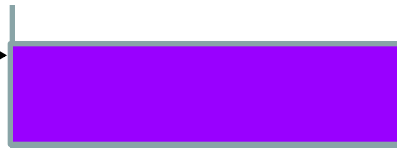
MTT



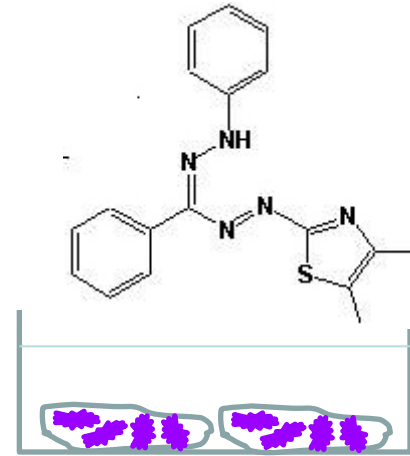
Live cells

Intact mitochondrion

Dissolving  
in DMSO



Mitochondrial  
succinate  
dehydrogenase



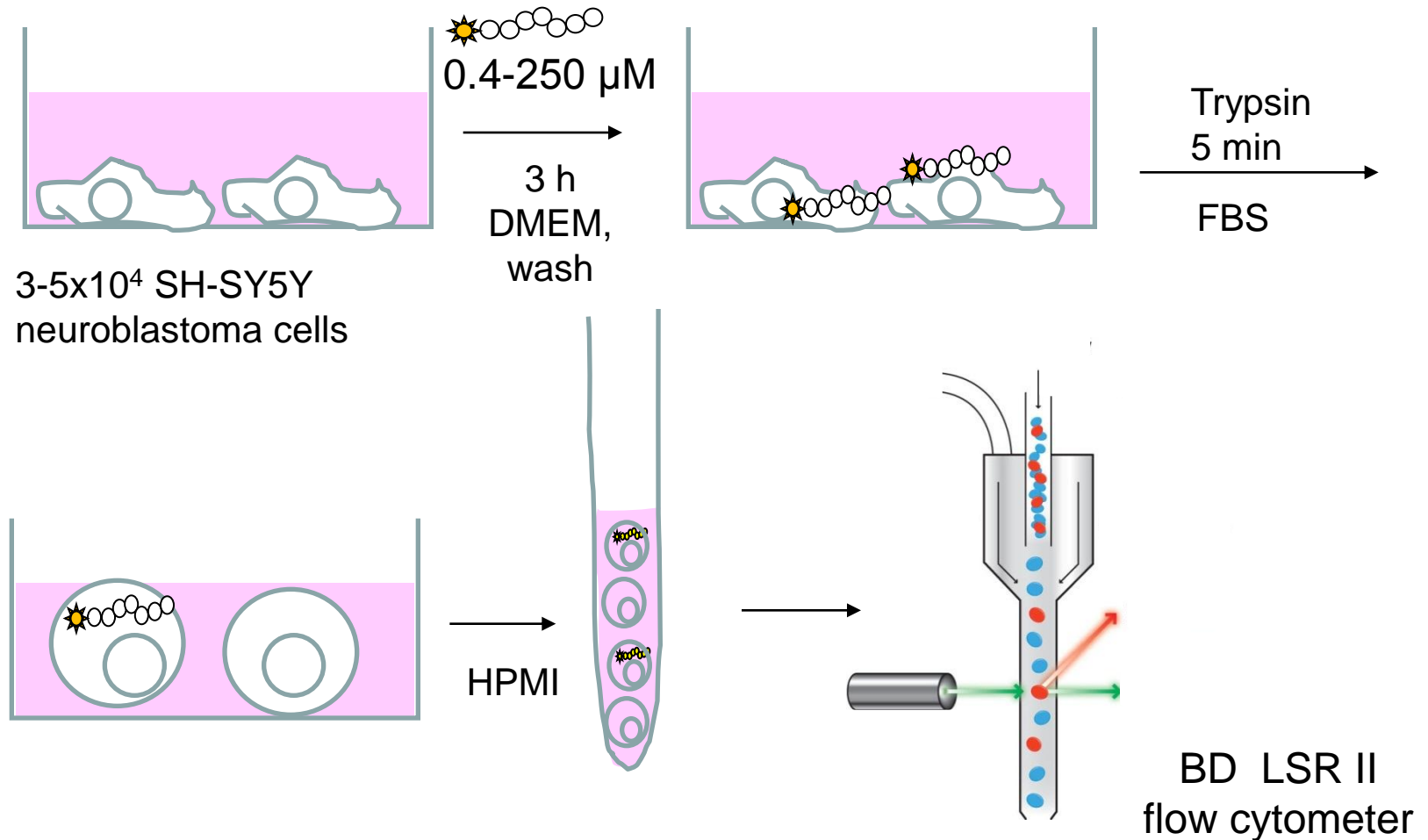
formazan

OD:  $\lambda_1=540$  nm; ref:  $\lambda_2=620$  nm

MTT: 3-[4,5-Dimethylthiazol-2-yl]-2,5-diphenyltetrazolium bromide

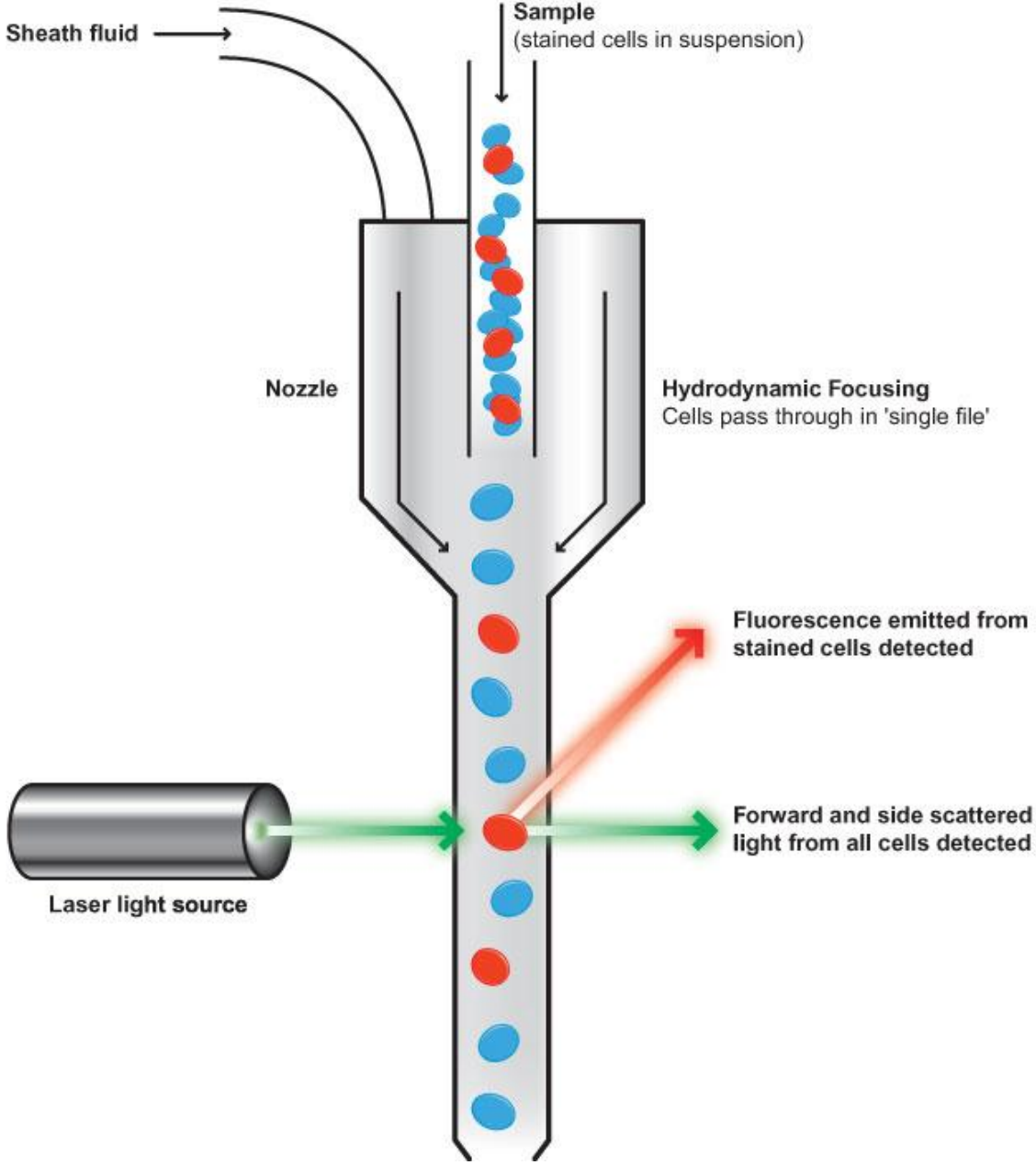
The peptides are not cytostatic in the conditions  
to be used for cellular uptake experiments

# Studying the *in vitro* cellular uptake of Cf-HSV peptides, flow cytometry

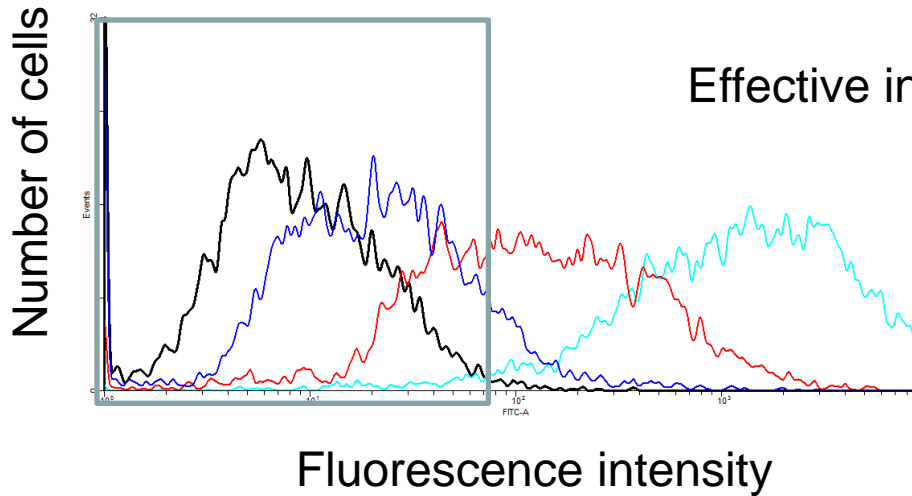


Coherent Sapphire laser, 22 mW,  $\lambda_{\text{exc}} = 488 \text{ nm}$   
FITC, LP 510, BP 530/30 (PE, LP 550, BP 576/26)

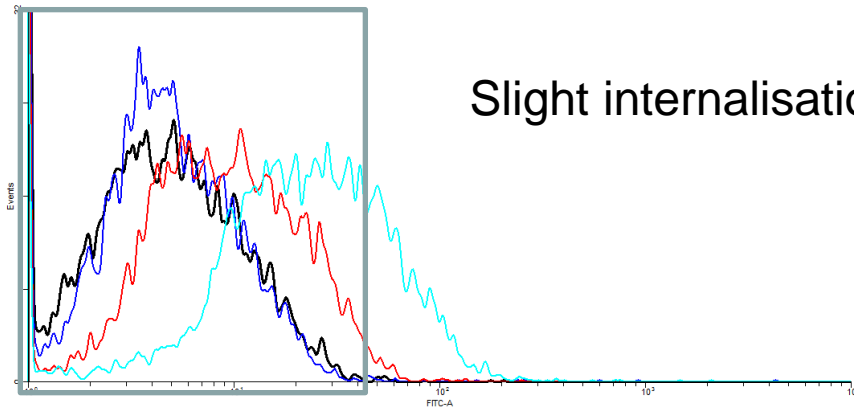
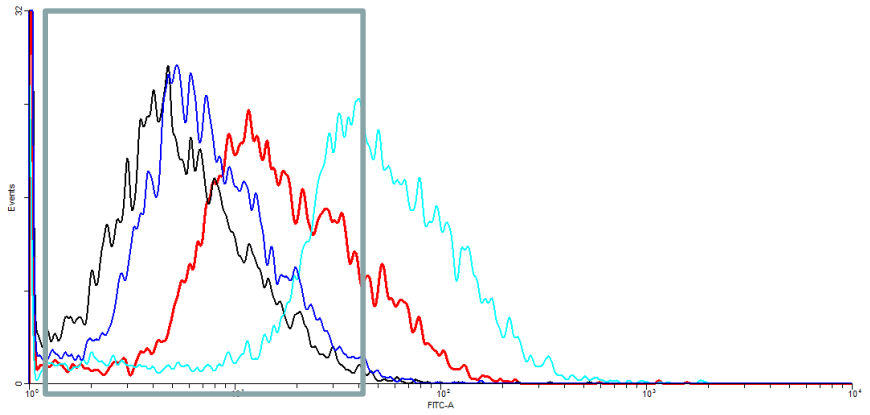
# Flow Cytometry



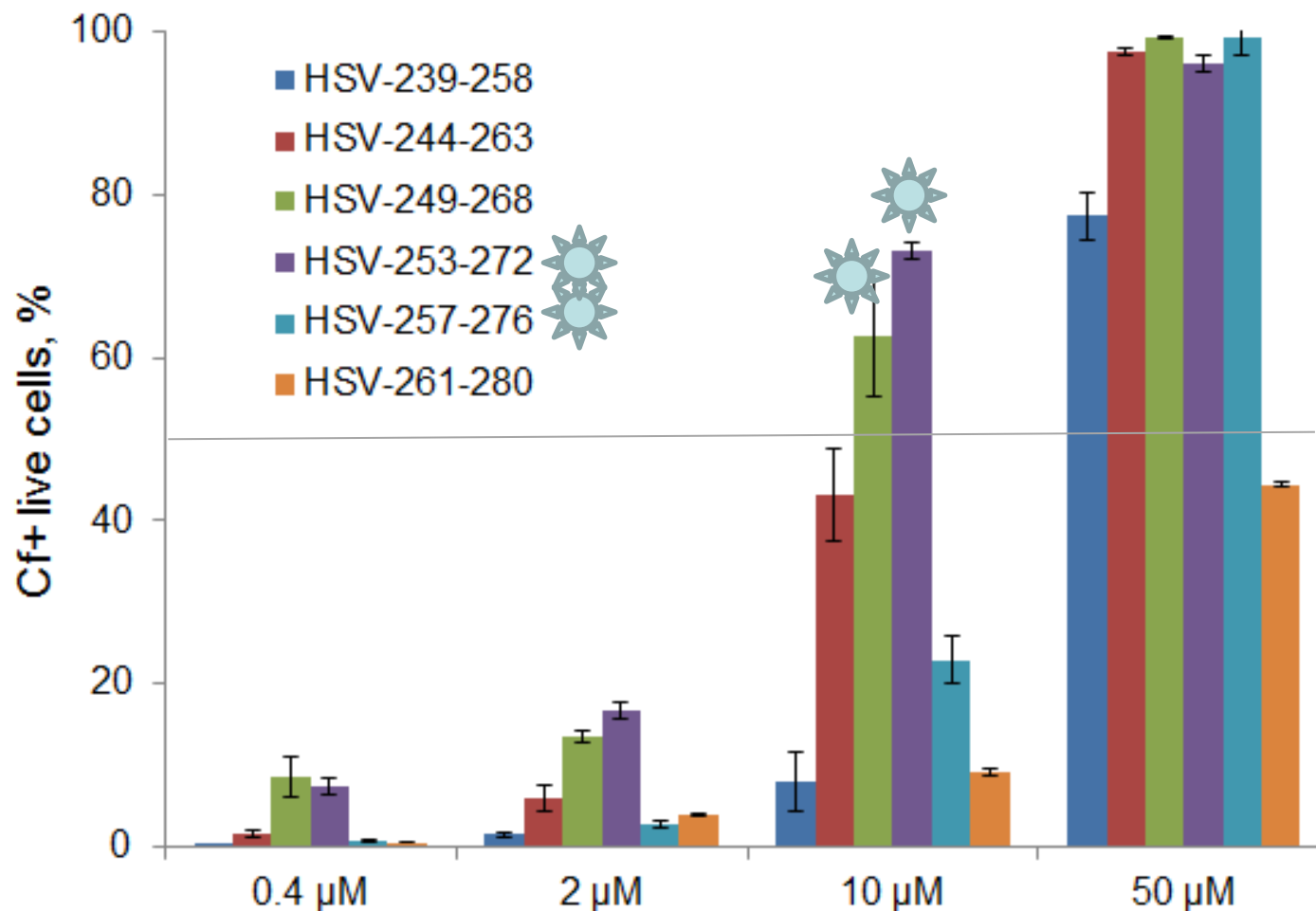
# Internalisation of Cf-HSV peptide into SH-SY5Y cells



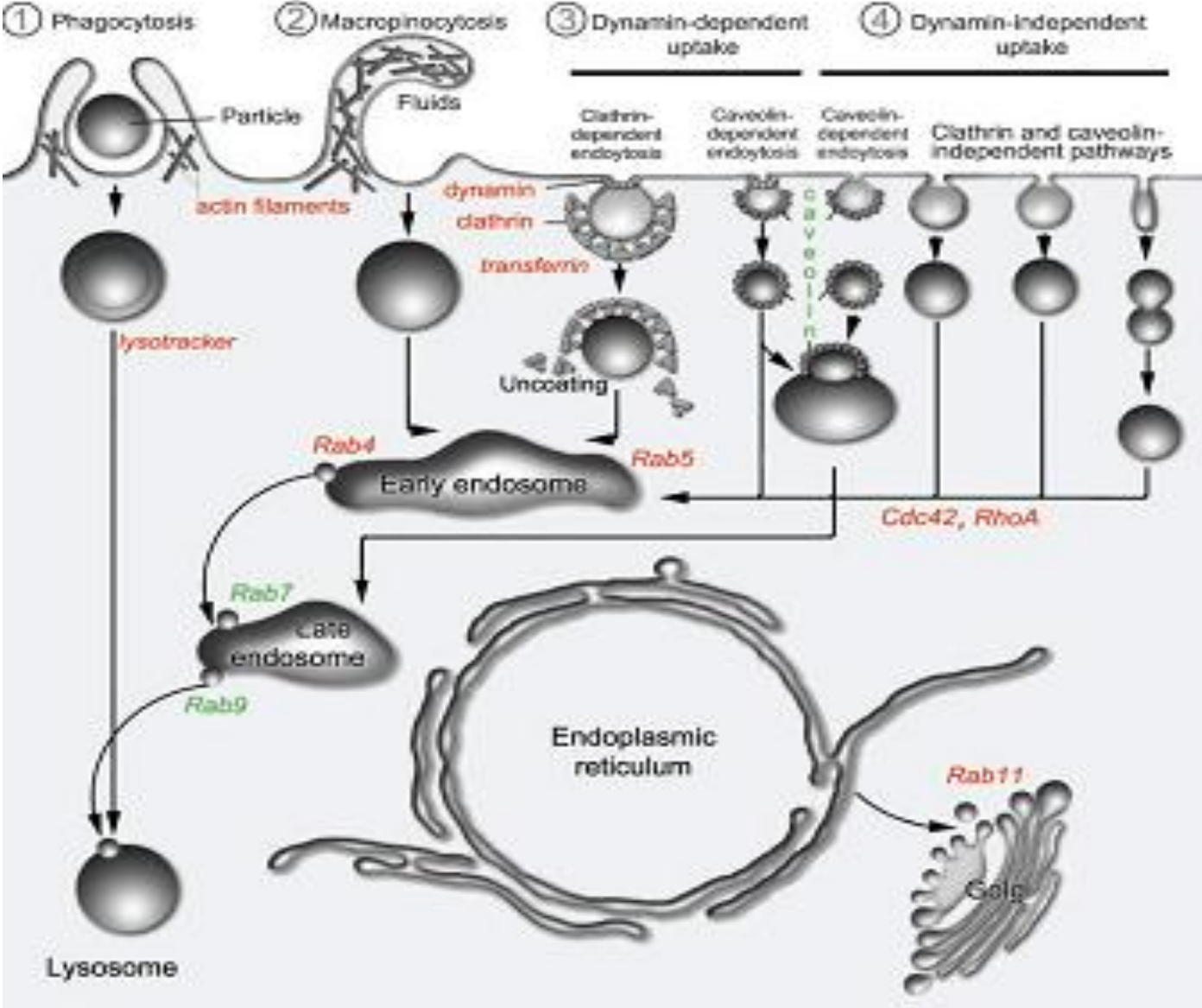
50  $\mu\text{M}$   
10  $\mu\text{M}$   
2  $\mu\text{M}$   
kontroll



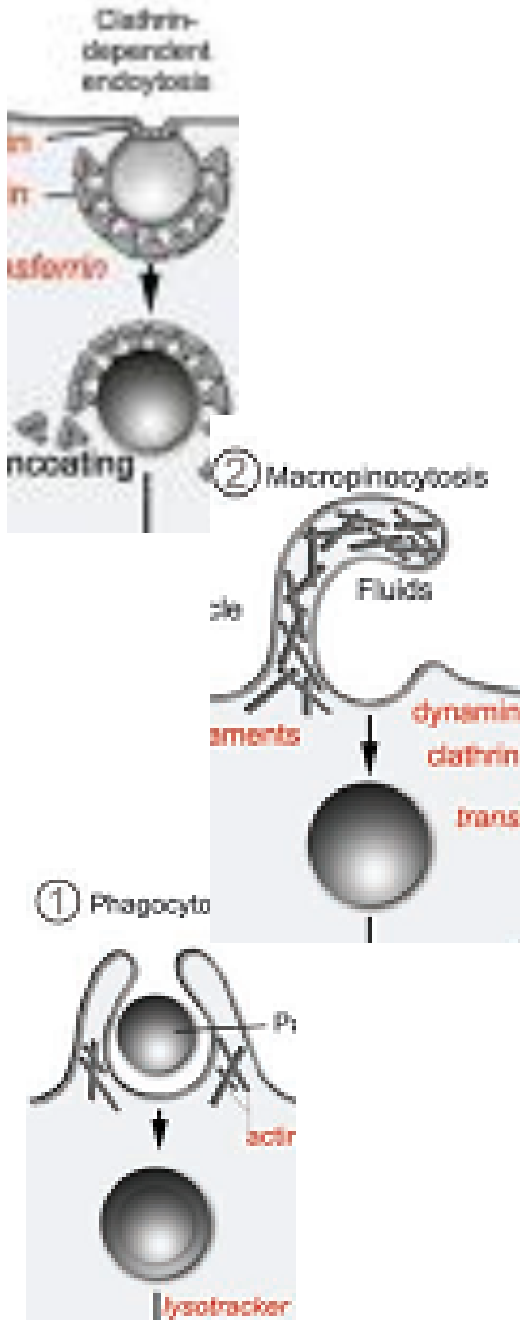
# Internalisation of Cf-HSV peptides into SH-SY5Y neuroblastoma cell



# Endocytosis pathways



# Inhibition of cellular uptake



Inhibitor	Mechanism	Type of endocytosis
Cytochalasin D	Actin polymerisation inhibitor	Macropinocytosis, clathrin dependent endocytosis <sup>1,2</sup>
EIPA (5-(N-ethyl-N-isopropyl)amylorid)	Selective Na <sup>+</sup> /H <sup>+</sup> antiport inhibitor	Macropinocytosis <sup>2,3</sup>
Colchicin	Microtubule polymerisation inhibitors	Pinocytosis <sup>4</sup>
Methyl-β-cyclodextrin	Cholesterol depletion from membrane	Caveola/lipid raft mediated endocytosis <sup>5</sup>

<sup>1</sup>Nakase et al, Mol. Ther. (2004) 10: 1011-1022

<sup>2</sup>Delwig et al, Arthr. Res Ther. (2006) 312: 1345-1360

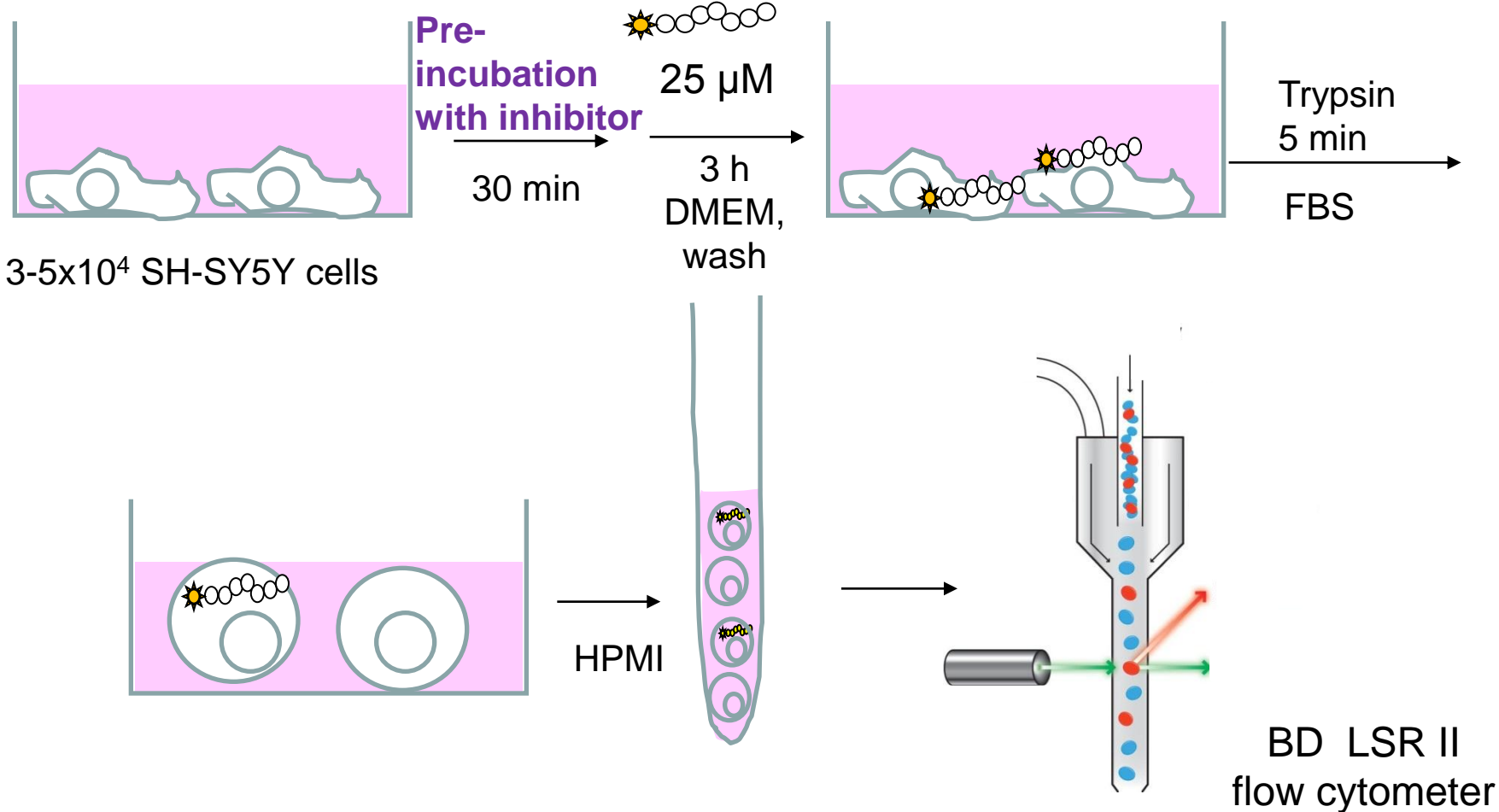
<sup>3</sup>Heikkilä et al, J. Virol. (2010) 84: 3666-3681

<sup>4</sup>Piasek et al, hematol. Blood Transf. (1985) 29: 511-513

<sup>5</sup>Rodal et al, Mol. Biol. Cell (1999) 10: 961-974

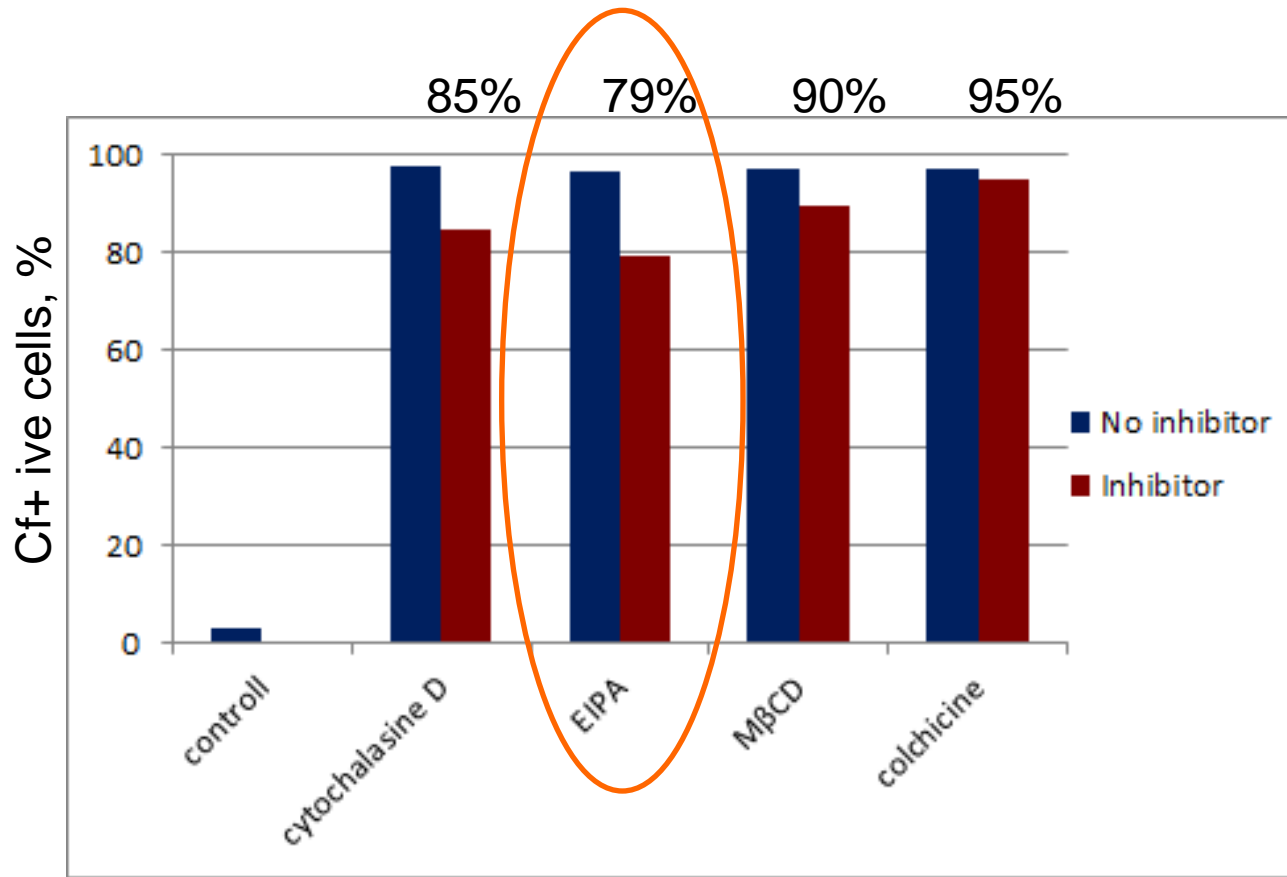


# Inhibition of the *in vitro* cellular uptake of a Cf-HSV gD peptide, flow cytometry



Coherent Sapphire laser, 22 mW,  $\lambda_{\text{exc}} = 488 \text{ nm}$   
FITC, LP 510, BP 530/30 (PE, LP 550, BP 576/26)

# Inhibition of the cellular uptake of a Cf-HSV gD peptide, flow cytometry

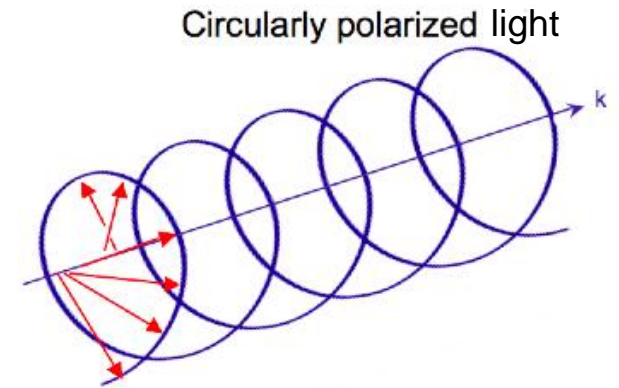
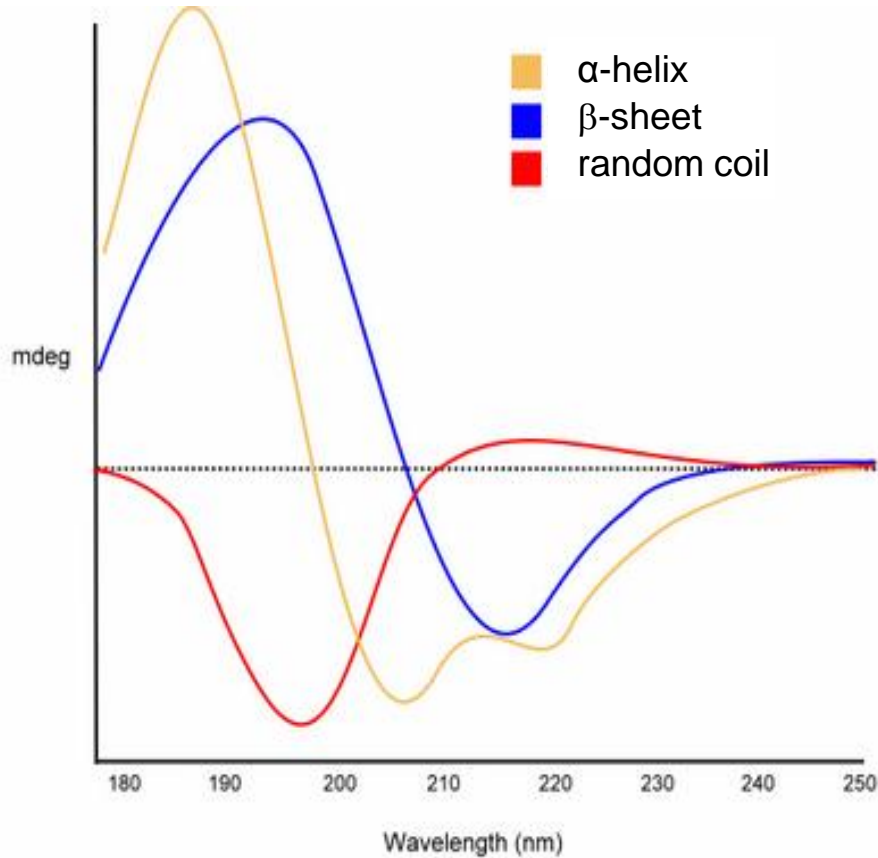


Macro-pinocytosis:  
compatible with  
**receptor mediated endocytosis**

Inhibitor concentration: 5 μg/ml 100 μM 2.5 mM 10 μM – 30 min preincubation

Peptide concentration: 25 μM

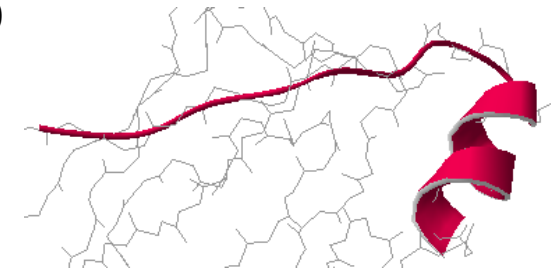
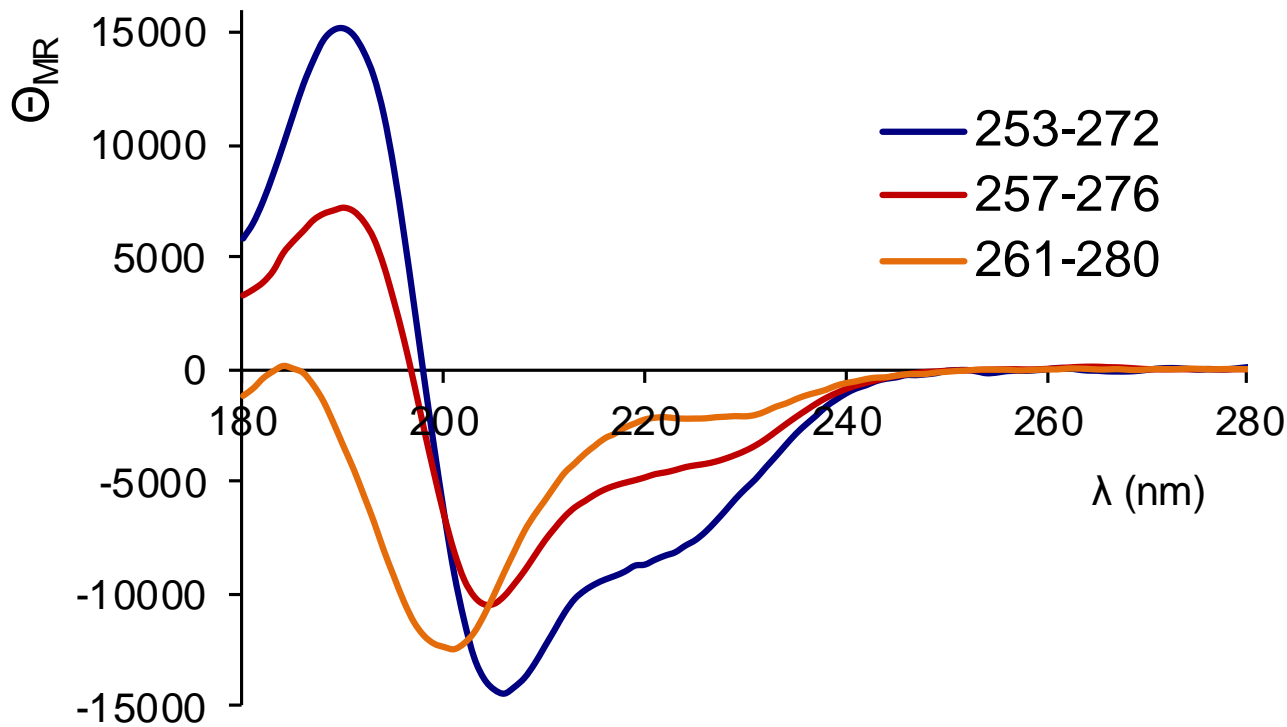
# Electronic Circular Dichroism spectra of different protein conformations



Optically active medium:  
 $\Delta\varepsilon = \varepsilon_L - \varepsilon_R$

# Secondary structure studies – ECD

## TFE – water (1-1 V/V)



Jasco 810 polarimeter  
c=0.5-0.7 mg/mL

Bősze, Zsila, Majer, Hudecz, Uray (2018) in: Proc. 35<sup>th</sup> European Peptide Symposium (Eds: Timmons, Hewage, Lebl), EPS, pp. 312-314

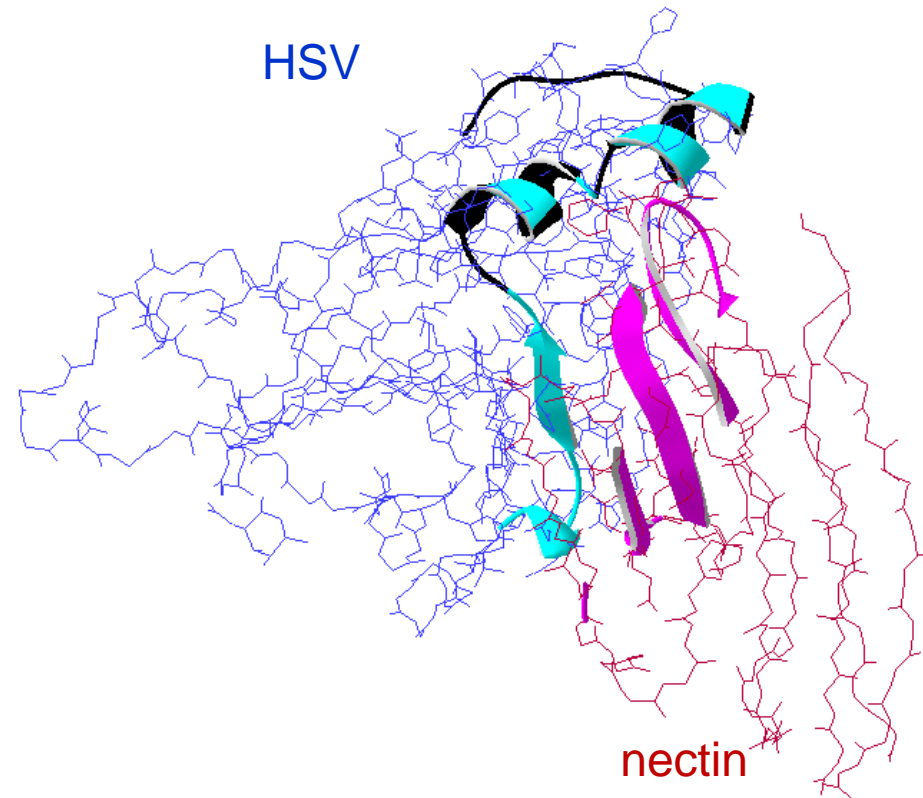
# Summary

Cf-labeled HSV-1 gD 20-mer (and 16mer) peptides internalised into SH-SY5Y neuroblastoma cells at different rates, in concentration dependent manner.

Some nectin-binding peptides internalised with high efficiency, even in small concentration

Uptake of peptide was inhibited by EIPA  
=> Method of entry: macropinocytosis  
– compatible with receptor mediated endocytosis

Peptides with pronounced  $\alpha$ -helical content internalised with higher efficiency



249 **IPENQRTVAVYSLKIAGWHGPKAP** 272

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Dr Ferenc Zsila



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*Thank you for your attention!*